

Matthew Bogyo

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EDUCATION

Massachusetts Institute of Technology, Cambridge, MA
Ph.D. in Biological Chemistry- Department of Chemistry, September 1997
Advisor: Dr. Hidde Ploegh

Bates College, Lewiston, ME
Bachelor of Science in Chemistry, May 1993
Advisor: Dr. Tom Wenzel

PROFESSIONAL AND ADVISORY POSITIONS

Positions and Employment

1991-93 Council on Undergraduate Research AIURP Fellow- Bates College, Lewiston, ME
1993-97 Graduate Student- Massachusetts Institute of Technology, Cambridge, MA.
1997-98 Post-Doctoral Fellow- Harvard Medical School, Boston, MA
1998-01 UCSF Faculty Fellow- University of California, San Francisco, San Francisco, CA.
2001-03 Group Leader, Head of Chemical Proteomics- Celera Genomics, South San Francisco, CA.
2001-03 Adjunct Faculty Member – UCSF Department of Pharmaceutical Chemistry
2003-09 Assistant Professor - Department of Pathology, Stanford University.
2004-09 Assistant Professor - Department of Microbiology and Immunology, Stanford University.
2005-09 Assistant Professor by courtesy–Department of Chemical and Systems Biology, Stanford University
2009-13 Associate Professor - Department of Pathology, Stanford University.
2009-13 Associate Professor - Department of Microbiology and Immunology, Stanford University.
2009-13 Associate Professor by courtesy–Department of Chemical and Systems Bio. ,Stanford University
2013- Professor - Department of Pathology, Stanford University.
2013- Professor - Department of Microbiology and Immunology, Stanford University.
2013- Professor by courtesy –Department of Chemical and Systems Biology, Stanford University

Other Experience and Professional Memberships

2000-01 Scientific Consultant- Rigel Pharmaceuticals, South San Francisco, CA.
2000-01 Scientific Consultant- Axys Pharmaceuticals, South San Francisco, CA.
2003-06 Scientific Consultant- Celera, South San Francisco, CA.
2003-10 Scientific Consultant- Proteolix, South San Francisco, CA.
2011-13 Scientific Consultant – Calithera Biosciences, South San Francisco, CA
2017-21 Scientific Consultant – Cortexyme, South San Francisco, CA
2016- Scientific Consultant – Vergent Biosciences
2018-21 Scientific Advisory Board – Cortexyme, Inc.
2010- Scientific Advisory Board – Benchfly
2008- Co-Founder and Member, Board of Directors – Akrotome Imaging Inc.
2017-21 Co-Founder and Head of Scientific Advisory Board – Facile Therapeutics
2011-15 Academic Editor – *PLoS One*
2015- Section Editor – *PLoS One*
2002-12 Editorial Board Member – *Biochemical Journal*.
2002- Editorial Board Member – *Chemistry and Biology*.
2002- Editorial Board Member – *Molecular & Cellular Proteomics*.
2005-09 Council Member – International Proteolysis Society
2005-07 Secretary - International Proteolysis Society
2007-09 President - International Proteolysis Society

2005-16 Faculty Member, Faculty of 1000
 2007-09 Member, DARPA funded Defense Science Study Group
 2007 Vice-Chair, Fifth International Proteolysis Society General Meeting, Petras, Greece
 2011 Co-Organizer, Seventh International Proteolysis Society General Meeting, San Diego, CA
 2009 Ad-Hoc Member – SBCB Study Section
 2011, 2012 Ad-Hoc Member – DDR Study Section
 2012, 2014 Ad-Hoc Member – EBIT study Section
 2015 Ad-Hoc Member – MSFE study Section
 2016- 2022 Standing Member – SBCA Study Section
 2013, 2015 Member of NCI Site Visit External Review Panel
 2009 NIH Peer Review – ARRA Challenge Grants
 2000- Member, American Chemical Society
 2003- Member of the American Society of Microbiologists
 2014 Organizer – Keystone Meeting – “The Chemistry and Biology of Cell Death” - Santa Fe, NM
 2014 Elected member - American Association of University Pathologists – Pluto Club
 2016 Co Chair – Imaging in 2020 Meeting – Jackson Hole, WI
 2016 Vice-chair – Gordon Research Conference – “Proteolytic Enzymes and Their Inhibitors”
 2018 Chair – Gordon Research Conference – “Proteolytic Enzymes and Their Inhibitors”

AWARDS AND FELLOWSHIPS

1991 Recipient of pre-doctoral fellowship from Council on Undergraduate Research
 1992 American Institute of Chemists award for outstanding performance in chemistry
 1992 American Chemical Society-Division of Polymer Chemistry award for organic synthesis
 1995 Recipient of MIT-Japan Science and Technology Prize
 2003 Recipient of Stanford University Terman Fellowship
 2004 Searle Scholar Award
 2005 Burroughs Wellcome Fund – Investigators in Pathogenesis of Infectious Disease Award
 2008 Strategic Program for Asthma Research – Early Excellence Award
 2012 Invited to attend NAS Kavli Frontiers of Science Symposium
 2014 Election to American Association of University Pathologists – Pluto Society

INVITED LECTURES

Claremont Colleges – Keck Sciences seminar speaker – Claremont, CA Feb 2024
11th Takeda Science Foundation Symposium – Invited speaker- Osaka Japan Jan 2024
UMass Amherst – Department of Chemistry Seminar Speaker – Amherst, MA Nov 2023
Tufts University – Department of Chemistry Seminar Speaker – Boston, MA Nov 2023
Ecole Polytechnique Federale de Lausanne – Dept of Biochemistry Seminar Lausanne Switzerland – Oct 2023
University of Edinburgh – Translational Chem. and Biomedical Imaging Seminar – Edinburgh, Scotland Oct 2023
Broad-MIT Seminar Series – Invited speaker. Boston, MA Oct 2023
Boston College – Department of Chemistry Seminar Series – Boston, MA Sept 2023
Tufts Medical School – Department of Microbiology Invited Speaker – Boston, MA Sept 2023
Activity Based Probes in Biology – Weizmann Institute, Rehovot Israel – July 2023
FASEB Meeting – The Optical Probes Conference: Discovery to Application – Speaker Palm Springs, CA May 2023
Molecular Imaging Symposium – World Molecular Imaging Society – Invited Speaker, Stanford, CA Jan 2023
EMBO Workshop – 20S Proteasome Degradation Pathway – Invited Speaker Rehovot, Israel Jan 2023
University of Florida – Department of Microbiology and Cell Science – Gainesville, FL Nov 2022
UCSF – CBI Chemical Biology Symposium – Invited Speaker San Francisco, CA Sept 2022
Imaging in 2020 Meeting – Invited Speaker – Grand Teton National Park, WY Sept. 2022
International Chemical Biology Society – West Coast Symposium – Invited speaker San Francisco, CA Sept 2022
Tri-Institutional Program in Chemical Biology Symposium – Invited Speaker – New York, NY Sept 2022
Gordon Research Conference – Proteolytic Enzymes and Their Inhibitors – Invited Speaker Italy, June 2022
World Molecular Engineering Network Meeting – Invited speaker – San Jose Del Cabo, Mexico May 2022
Texas A&M Univeristy – Department of Chemistry – Invited Speaker (virtual) April 2022
Annual Symposium of the Institute of Chemical Immunology – Keynote Speaker Leiden. Netherlands Mar 2022
Boston Protein Degradation Meeting – Invited Speaker Boston, MA Mar 2022
Norwegian Biochemical Society National Meeting - Invited Speaker Feb12, 2022 Tromso, Norway

SPIE Photonics West National Meeting – Invited Speaker Jan. 22, 2022 San Francisco, CA
16th Annual Peptide Therapeutics Foundation Symposium – Invited Speaker Oct. 22, 2021 San Diego, CA
University of Wisconsin-Madison– Department of Biochemistry – Invited Speaker Oct. 11, 2021 Madison, WI
RAS Targeted Drug Development Summit – Virtual Meeting – Invited Speaker – Sept. 21, 2021
University of Southern California – Department of Chemistry – Invited Speaker – Aug. 9, 2021 Los Angeles, CA
University of Edinburgh - UoE Centre for Inflammation Research – Invited speaker-Virtual-May 18, 2021
Washington University – Radiology Seminar Series – Invited Speaker – Virtual – Feb. 23, 2021
Wayne State University – Dept. of Biochemistry, Microbio and Immunology – Invited lecture – virtual Oct 20, 2020
Nature Conferences – Translational Chemical Biology – virtual conference – invited speaker Oct. 5, 2020
Merck Research Labs – Invited Speaker – Zoom Lecture – March 28, 2020
North American Protein Degradation Congress – Invited speaker. San Diego, CA Feb 2020.
University of Florida – Department of Medicinal Chemistry – CNPD3 Seminar Speaker, Gainesville, FL Nov 2019.
The Scripps Research Institute – Invited speaker – Jupiter, FL Nov. 2019.
University of California, Irvine – Department of Chemistry Invited speaker. Irvine, CA Nov. 2019
11th General Meeting of the International Proteolysis Society – Invited Speaker, Prague, Czechia Oct. 2019
7th Modern Solid Phase Peptide Synthesis & Its Applications Symposium – Invited Speaker – Palm Cove, Australia – Sept 2019.
FASEB – Protein Lipidation Conference – Keynote – Richard Gibbs Lecture, Olean, NY – July 2019
World Molecular Engineering Network Meeting – Invited speaker, San Jose del Cabo Mexico May 2019
Baylor University – Department of Chemistry and Biochemistry - Invited Speaker May 2019
HHMI, Janelia Research Campus – “Chemical Tools in Biology” – Invited speaker – May 2019
ABPP 2019 Target Discovery and Visualization – Keynote lecture – Leuven Belgium March 2019
Columbia University – Department of Microbiology – Invited speaker – Feb 2019
University of Pennsylvania – Department of Microbiology and Immunology – Invited speaker Feb 2019
Ben-Gurion University – Chemical Biology Symposium – Invited Speaker, Israel Jan 2019
University of Chicago – Department of Chemistry Seminar Series, Chicago, IL Nov 2018
University of Virginia – Department of Chemistry Seminar Series, Charlottesville, VA Nov 2018
HHMI Janelia Research Seminars – “Probe Fest 2018”, Janelia Campus, VA Oct 2018
World Molecular Imaging Society Conference – Invited Speaker Seattle, WA Sept 2018
Queenstown Molecular Biology Meeting – Invited Speaker – Queenstown, New Zealand Aug 2018
University at Buffalo – Department of Chemistry Seminar Speaker – Buffalo, NY Apr 2018
University of Michigan – Department of Medicinal Chemistry Seminar Speaker – Ann Arbor MI Apr 2018
Chemical Probes in Biology Symposium – Invited Keynote Speaker – Oxford UK Mar 2018
Academy of Science of the Czech Republic – Invited Speaker, Prague Czech Republic Mar 2018
GTCBio – Protease Inhibitors in Drug Discovery – Keynote Lecture, San Diego, CA Feb 2018
University of Texas Southwestern – Dept of Biochemistry Seminar Series Speaker – Dallas TX Jan 2018
Oregon Health and Sciences University – Chemical Biology Symposium – Speaker – Portland OR Dec 2017
Memorial Sloane Kettering Cancer Institute – Invited speaker “Intraoperative Imaging Technologies”– Dec 2017
University of Illinois – Department of Chemistry Seminar Series Speaker – Urbana IL Dec 2017
Genomics Institute of the Novartis Research Foundation – Invited Speaker – San Diego CA Nov 2017
Hans Fischer Symposium – Invited Speaker, Munich Germany Nov 2017
University of Pittsburgh – Department of Chemistry Seminar Series Speaker – Pittsburgh PA Oct 2017
IUPAC International Symposium on Bioorganic Chemistry – Invited Speaker, Konstanz Germany, Sept 2017
University of Missouri – Department of Chemistry Seminar Series Speaker – Columbia MO Sept 2017
World Molecular Imaging Conference – Invited speaker – Philadelphia PA Sept 2017
Georgia State University – Dept. of Chemistry- seminar series speaker – Atlanta GA Aug 2017
HHMI, Janelia Research Campus – “Chemical Tools in Biology” – Invited speaker – Ashburn, VA April 2017
Vanderbilt University – Vanderbilt Institute for Chemical Biology Seminar Series – Nashville, TN Feb 2017
SPIE Photonics West– Molecular Guided Surgery Session – Invited speaker- San Francisco – Jan 2017
Duke Cancer Center - Radiation Oncology & Imaging Program – Invited Speaker – Jan 2017
1st international PSL* Chemical Biology Symposium – Invited Speaker – Paris France –Dec 2017
UC Berkeley – Structural & Quantitative Biology Seminar Series Speaker – Berkeley, CA – Nov 2017
B-DEBATE Workshop – Biomedical Imaging – Invited Speaker – Barcelona, Spain – Nov 2016
H3D Symposium 2016 – Invited Speaker – Malaria and Tb drug discovery– South Africa – Nov 2016
Memorial Sloane Kettering Cancer Institute – Invited speaker –“Intraoperative Imaging Technologies”– Oct 2016
Johnson Symposium – Invited Speaker – Department of Chemistry, Stanford University Oct 2016
UMass Amherst – Department of Chemistry – Invited speaker- Amherst, MA Oct 2016
Imaging in 2020 – “Molecular Imaging for Diagnosis and Surgery/Therapy” – Jackson Hole, WY Sept 2016
2nd International Symposium – ‘Protease World in Health and Disease’ – Kiel Germany Sept 2016
World Molecular Imaging Conference – Optical Surgical Navigation Symposium – New York, NY Sept 2016
Janssen Pharmaceuticals – Invited speaker – San Diego, CA Aug 2016

Gordon Conference – Proteolytic Enzymes and Their Inhibitors – invited speaker June 2016
Memorial Sloane Kettering Cancer Institute – Invited speaker – Department of Radiology – May 2016
Hamline University – 17th 3M/Ronald A. Mitsch Lecture – April 2016
University of British Columbia – Center for Blood Research – Vancouver, Canada Mar 2016
University of Rochester – Department of Chemistry Seminar series – Mar 2016
Keystone Meeting – Drug Discovery for Parasitic Diseases – Lake Tahoe, CA Jan 2016
Pacificchem 2015 – National Meeting – Symposium Organizer and speaker, Hawaii, Dec 2015
Eli Lilly Inc. – Invited Speaker – Indianapolis, ID Nov 2015
University of Kansas – Training grant symposium – Keynote Speaker – Nov 2015
The Beijing Symposium 2015 - "Frontiers at the Chemistry and Biology Interface" Beijing China Oct 2015
14th International Conference on the Chemistry of Antibiotics and Other Bioactive Substances – Keynote Speaker – Galveston, TX Oct 2015
International Proteolysis Society – 9th General Meeting – Penang Malaysia Oct 2015
Agensys Inc. – Invited Lecture – Santa Monica, CA Sept 2015
Harvard University – Department of Microbiology and Immunology Seminar Series – Boston, MA Sept 2015
Genentech – Invited Lecture – South San Francisco, CA Sept 2015
World Molecular Imaging Conference – Optical Surgical Navigation Symposium – Hawaii Sept 2015
American Chemical Society – National Meeting – Symposium Organizer and speaker, Boston, MA Aug 2015
20th American Peptide Symposium – Invited Lecture – Orlando, FL June 2015
Gordon Conference – High Throughput Chemistry and Chemical Biology – New London, NH June 2015
Gordon Conference – Bioorganic Chemistry – Andover, NH June 2015
University of Minnesota - 15th Annual NIH Chemistry-Biology Interface Symposium – May 2015
University of Georgia – Department of Cellular Biology – Seminar series, Athens, GA April 2015
Pluto Club Meeting – New Member Induction Lecture – Playa del Carmen, Mexico March 2015
GTCBio – Protease Inhibitors in Drug Discovery – Keynote Lecture, San Diego, CA Feb 2015
University Washington – Department of Global Health, Pathobiology Seminar Series – Seattle, WA Feb 2015
Washington University – Department of Biochemistry and Molecular Biophysics, St. Louis, MO Dec 2014
University of Wroclaw – Department of Chemistry, Invited Speaker, Wroclaw Poland Oct 2014
Brazilian Symposium on Chemistry and Physiology of Proteases and their Inhibitors – Invited speaker. Sao Carlos Brazil, Sept 2014
University of Montpellier – Invited Speaker, Montpellier France Sept 2014
28th Protein Society annual Symposium – Invited speaker, San Diego, CA July 2014
Gordon Conference – Proteolytic Enzymes and Their Inhibitors – invited speaker June 2014
National ACS of Medicinal Chemistry Meeting – "Novel Chemistries Keynote" Invited Speaker May 2012
24nd World Molecular Engineering Network Conference – Cabo San Lucas Invited Speaker May 2014
Northeastern University – Department of Chemistry Spring colloquium Series – Invited Speaker – April 2014
University of Southern California – Cellular Homeostasis Lecture Series – Invited Speaker- March 2014
Chemprobe: COST meeting – Cambridge, UK – Keynote speaker. March 2014
Keystone Meeting – "The Chemistry and Biology of Cell Death" – Invited Speaker and Organizer – Feb 2014
University of California, Berkeley – Infectious Diseases & Immunity/CEND – Invited speaker – Nov. 2013
University of California, San Francisco – Division of Experimental Medicine – Invited Speaker – Nov 2013
8th General Meeting of the International Proteolysis Society – Invited Speaker- Cape Town S. Africa – Oct 2013
University of Utah – Department of Biochemistry – Invited speaker – Oct 2013
University of Vermont – Department of Microbiology Retreat – Keynote speaker – Aug 2013
12th International Congress on Toxoplasmosis - Invited speaker, Oxford UK.
Genentech- Seminar and consulting trip – protein science group – June 2013
Biogen/Idc – Seminar and site visit – April 2013
245th American Chemical Society National Meeting– "Chemical Biology of Infectious Diseases" symposium - New Orleans - April 2013
University of South Florida – Department of Chemistry Seminar – March 2013
University of Wisconsin – Department of Chemistry Seminar – March 2013
University of Washington – Department of Chemistry Seminar – Feb 2013
Oregon Health and Science University – Department of Cell & Developmental Biology seminar – Feb 2013
Bowdoin College – Department of Chemistry Seminar – Feb 2013
Bates College – Department of Chemistry Seminar – Feb 2013
AKIRA Project –symposium "Towards Understanding of Immune Dynamism" Osaka, Japan Oct 2012
Gordon Conference – Biology of Host-Parasite Interactions – invited speaker June 2012
Gordon Conference – Proteolytic Enzymes and Their Inhibitors – invited speaker June 2012
Korean Society for Biochemistry and Molecular Biology – General Meeting – invited speaker May 2012
22nd World Molecular Engineering Network Conference – Cabo San Lucas Invited Speaker May 2012
University of Minnesota – Department of Chemistry Seminar Series – invited speaker April 2012

American Chemical Society National Meeting – “Proteases: from Mechanism to Drug Development” Mar 2012
Harvard School of Public Health - “Cellular and Molecular Biology of Parasites” course lecturer – Mar 2012
Harvard University – Department of Chemistry – Invited Speaker Mar 2012
University of Vermont – Department of Microbiology and Molecular Genetics Seminar Series – Mar 2012
Case Western University – Cancer Center Seminar Series – Invited Speaker Mar 2012
Gordon Conference – Plasminogen Activation and Extracellular Proteolysis – invited speaker Feb 2012
University of Michigan – Department of Microbiology and Immunology Seminar Series – Feb 2012
Hoffmann La Roche, Basel Switzerland – Chemistry Frontier Forum – invited lecturer – Jan 2012
National Cancer Institute – Molecular Discovery Program Seminar series – invited lecturer – Nov 2011
University of Lausanne – Lecturer for “Proteases in Health and Disease” course – Nov. 2011
University of California, Berkeley – Infectious Diseases & Microbial Pathogenesis Seminar Series – Sept. 2011
University of Queensland – Institute for Molecular Biosciences – Invited Speaker – Sept. 2011
Queensland University of Technology – Inst. of Health and Biomed. Innovation – Invited Speaker – Sept 2011
Nankai University – College of Life Science, Invited Speaker, Tianjin, China June, 2011
Canadian National Proteomics Network Symposium – Keynote Speaker, Banff, Canada, May 2011
Frontiers in Chemical Biology Symposium – Invited speaker, Shenzhen, China, April 2011
SRI international – Biosciences Division, Invited Speaker, March, 2011
Tel Aviv University – Imaging workshop, Invited Speaker, March, 2011
Hebrew University – Department of Pharmacology Seminar Series, March 2011
Yale University – Chemical Biology Seminar Series, Invited Speaker, March 2011
Boston College – Department of Chemistry Seminar Series, Invited Speaker, Feb, 2010
PacifiChem 2010 - Molecular Probes and Fluorophores for Cellular Imaging. Invited speaker, Hawaii Dec. 2010
Laboratory Medicine Day Theme: Malaria - University of Lund, Lund Sweden, Nov. 2010
EMBO Conference Series – Chemical Biology 2010 – Invited Speaker, Sept. 2010
XIIth Symposium on Proteases, Inhibitors and Biological Control – Portoroz Slovenia Sept, 2010
EFMC-ISMC Symposium on Medicinal Chemistry – Invited Speaker, Brussels, Belgium, Sept. 2010
University of California Santa Cruz – Dept. of Chemistry retreat – Keynote Speaker, Sept. 2010
American Chemical Society - Division of Biological Chemistry meeting Boston, MA Aug 2010
Gordon Conference – Cell Death, Invited Speaker Newport RI, Aug 2010
Netherlands Cancer Institute – Invited Speaker – Amsterdam, Holland July 2010.
IMP Symposium - Seeing more: Emerging Technologies in Microscopy – Vienna Austria May 2010
US HUPO Annual Conference – Invited Speaker Denver, CO March 2010.
Scripps Research Institute – Structure & Chemistry Affinity Group lecture series – Feb 2010
Johns Hopkins University – School of Public Health Lecture series – Feb 2010
Vanderbilt University – Institute of Chemical Biology seminar series – Dec 2009
Novartis Lecture Series – Invited Speaker, Basel Switzerland Nov 2009
Ubiquitin Drug Discovery Meeting – Invited Speaker, Philadelphia, PA Oct 2009
MalERA Meeting – 1 of 25 Invited participants, Harvard Medical School Oct 2009
Nature Publishing Group SciCafe – Invited Speaker, San Francisco, CA Sept 2009
Imaging in 2020 – Molecular imaging meeting – invited speaker, Jackson, WY Sept 2009
Korean Society for Microbiology – International Symposium, Daejeon, Korea, June 2009.
FASEB Meetings - Proteases in Hemostasis and Vascular Biology, Carefree, Az, June 2009.
Abbott Research Labs – Seminar series, Abbott Park, IL – April 2009
University of Kentucky – College of Pharmacy Seminar Series – April 2009.
Northwestern University School Medicine –Gastroenterology Division Lecture Series – March 2009.
Mt. Sinai School of Medicine – Department of Microbiology and Immunology Seminar Series – Dec 2008.
National Institutes of Health – Chemistry seminar series Bethesda, MD November 2008
Gordon Conference – Proteolytic enzymes and their inhibitors - New Hampshire – July 2008
Gordon Conference – Bioorganic Chemistry - New Hampshire – June 2008
Department of Molecular Biology – University of Texas Southwestern Medical Center – Dallas Texas April 2008
Max-Planck-Institut für Züchtungsforschung – Department Seminar – Cologne, Germany – March 2007.
Genomes to Systems Conference - Chemical Genomics: Small molecules with large effects – Manchester, UK March 2007.
Lorne Conference on Protein Structure and Function – Lorne, Victoria, Australia February 2008
Department of Microbiology & Immunology – Albert Einstein College of Medicine – seminar series – Dec 2007
Cambridge Healthtech Institute – in vitro molecular imaging meeting – San Diego November 2008
International Proteolysis Society – General meeting – invited speaker – Petras, Greece Oct. 2007
Human Proteome Organization (HUPO) – 6th Annual World Congress – Seoul, Korea Oct. 2007
Drug Action and Chemical Biology in the Post-Genomic Era – Invited speaker – Vienna, Austria Aug 2007
Societat Catalana de Biologia - Perspectives in Genomics and Proteomics – Barcelona, Spain Jul. 2007
American Peptide Symposium – 20th National Symposium – Montreal, Canada June 2007

Advances in Optics for Biotechnology, Medicine, and Surgery – Invited Speaker – Naples, FL June 2007
Seattle Biomedical Research Institute - Seattle Parasitology Conference - Seattle, WA – May 2007
University of California Riverside – Dept. of Cell Biology and Neuroscience seminar – Riverside, CA – May 2007
Searle Scholars – Lecture at annual meeting – Chicago, IL April 2007
Molecular Cell Biology and Biotechnology seminar series – Virginia Tech – Feb 2007
Department of Pharmacology Seminar – Yale University – Feb 2007
Biochemical Society – Proteomics and Proteolysis – London England Jan 2007
Department of Chemistry Seminar - University of California, Berkeley – Nov 2006.
Department of Microbiology Seminar – University of Vermont – Oct 2006
American Chemical Society Southwest Regional Meeting – Invited lecture – Oct 2006
5th International Conference on Cysteine Proteinases and Their Inhibitors: From structure to regulation and biology – Portoroz, Slovenia Sept. 2006
18th International Congress on Fibrinolysis and Proteolysis – San Diego, CA Aug 2006
Gordon Research Conference – Proprotein Processing, Trafficking & Secretion- July 2006
Novartis Institutes for Biomedical Research – Weekly seminar – Cambridge, MA May 2006
University of Iowa – Bioinformatics lecture series – April 2006
Association of Biomolecular Resource Facilities – Annual Meeting – Long Beach, CA Feb 2006
Toxoplasma Infections in the Immune Competent Host-Possible Application to Human Neuropsychiatric Diseases – Annapolis, MD November 2005
University of Southern California – Chemical Biology Seminar Series – November 2005
University of California San Francisco, Chemistry and Chemical Biology seminar series – Oct 2005
International Proteolysis Society – 4th general meeting – Quebec, Canada Oct 2005
Society for Molecular Imaging – 4th Annual Meeting – Cologne, Germany Sept 2005
University of California San Francisco –Biophysics/Chemistry & Chemical Biology Seminar Series– Oct 2005
IXth International Symposium on Proteinase Inhibitors and Biological Control – Brdo, Slovenia June 2005
Celera Genomics – Chemistry Department Seminar Series – June 2005
American Association for the Study of Liver Diseases - Functional Genomics and Proteomics of Liver in Health and Diseases – June 2005
Genentech – Invited by Dr. Robert Lazarus Department of Protein Engineering - March 2005
Washington University School of Medicine - Department of Molecular Microbiology Seminar - March 2005
Scripps Research Institute – Molecular and Cell Biology Affinity Group seminar series – November 29, 2004
American Society of Microbiologists - Conference on Functional Genomics and Bioinformatics Approaches to Infectious Disease Research – October 2004
Molecular Parasitology Meeting XV – Woods Hole, MA – September 2004.
American Chemical Society – National Meeting. “Genomic Approaches to Enzymology” – August 2004
Gordon Research Conference – Proteolytic Enzymes and Their Inhibitors - July 2004
McGill University - Chemical Biology/Archibald Macallum Seminar Series – June 2004
University of Washington School of Medicine–Department of Genome Sciences Lecture Series – June 2004
Lorne Conference on Protein Structure and Function – Lorne, Victoria, Australia February 2004
Molecular Approaches to Malaria - Lorne, Victoria, Australia February 2004
American Society for Cell Biology National Meeting – Special Session “The interface Between Small Molecule Chemistry and Cell Biology”– December 2003
International Proteolysis Society - 3rd general meeting - Nagoya, Japan November 2003
Oregon Health & Sciences University - Keynote Lecture for Department of Microbiology and Immunology Retreat. November 2003
Horizon Symposium - Third Symposium on Proteolysis - Verona, Italy October, 2003
Genomics on Target - IBC conference Boston, MA October , 2003
Proteomics: The Chemical Tools and Challenges - Churchill College, Cambridge, UK July 2003
University of Leiden - Department of Chemistry Seminar Series - May 2003
University of Pennsylvania Medical School - George Raizzis seminar series November 2002
American Association for Cancer Research – “Ubiquitination in Normal and Cancer Cells” Vancouver, British Columbia. November 2002
American Association for Cancer Research – “Proteases, Extracellular Matrix, and Cancer” Hilton Head, South Carolina October 2002
International Society for Fibrinolysis and Proteolysis –Munich, Germany September 2002
American Chemical Society- National Meeting Boston, Massachusetts August 2002
Gordon Research Conference- “Proteolytic enzymes and their inhibitors” July 2002
ASBMB Annual Meeting- Experimental Biology 2002, Proteomics section April 2002
University of British Columbia - Protein Engineering Network Symposium - March 2002
Gordon Research Conference- “Chemistry and Biology of Peptides” February 2002
International Proteolysis Society - 2nd general meeting - Munich, Germany November 2001

Israeli Society for Combinatorial Technologies- Israeli CombiTech Symposium Rehovot, Israel October 2001
American Chemical Society- National Meeting Chicago, Illinois August 2001
5th International Symposium on Mass Spectrometry in the Health and Life Sciences- "Molecular and Cellular Proteomics" San Francisco, California August 2001
Merck Research Laboratories - Rahway, New Jersey July 2001
7th Brdo Symposium on Proteinase Inhibitors and Biological Control -Brdo Slovenia June 2001
Cold Spring Harbor Symposium- "Proteolysis and Biological Control" May 2001
University of Munich- Department of Clinical Biochemistry Munich, Germany February 2001
Albert-Ludwigs-University Freiburg- Institute for Molecular Medicine Freiburg, Germany February 2001
Surface Logix- Boston, Massachusetts February 2001
Affimax Research Institute- Santa Clara, California February 2001
The Burnham Institute- La Jolla, California January 2001
Axys Pharmaceuticals - South San Francisco, California November 2000
Rigel Pharmaceuticals - South San Francisco, California November 2000
Cysteine Proteinases and their Inhibitors: The new Millennium- Portoroz, Slovenia September 2000
Conference of the World Molecular Engineering Network- Cabo San Jose, Mexico April 2000
Axys Pharmaceuticals - South San Francisco, California March 2000
University of Utah- Department of Biochemistry March 2000
The Scripps Research Institute- November 1999
American Chemical Society- Northwest Regional Meeting June 1999
University of California, San Francisco-Department of Pharmaceutical Chemistry retreat December 1998
Gordon Research Conference- "Proteolytic enzymes and their inhibitors" June 1998
Janssen-Cilag Club de la Transplantation meeting - Paris, France October 1997
American Peptide Symposium – Nashville, Tennessee June 1997
Cold Spring Harbor Symposium- "Biology of Proteolysis" April 1997
University of California, San Diego - Department of Pathology March 1997

TEACHING

Graduate Programs

Cancer Biology
 Microbiology and Immunology
 Chemical and Systems Biology

Training Grants

Cancer Biology
 Microbiology and Immunology
 Chemical and Systems Biology
 Molecular Pharmacology Training Program

CSB 220 – Co-Founder of course with T. Wandless, D. Hershlag, J. Chen "Chemistry of Biological Processes". Primary Lecturer Fall 2004, Fall 2005, Fall 2007
CSB240 – Lecturer Winter 2009, 2010, 2011
CSB260 – Lecturer and discussion leader – Spring 2007, 2008
BioE222B – Lecturer Winter 2010-Current
Bio 241 – Discussion leader Winter 2004, Lecturer Winter 2006, Discussion leader Winter 2006
CBio 241 – Lecturer Fall 2003- Current Discussion leader Fall 2005
CBio 242 – Lecturer Summer 2007-Current
CBio 280 – Discussion leader Spring 2005
M&I 210 - Lecturer Winter 2005 - Current
M&I 215 – Lecturer Winter 2005 - Current
BMI234 – Lecturer Winter 2005, 2007

PATENTS

1. H. L. Ploegh, H. A. Chapman, **M. Bogyo**, and P.R. Wolf. "Suppression of Immune Responses Via Inhibition of Cathepsin S". US 7,285,525 B2 issued Oct. 23, 2007.
2. **M. Bogyo**, S.H.L. Verhelst and A.M. Sadaghiani "Design and Synthesis of Novel Cysteine Protease Inhibitors" US Utility Filing 11/762,735.
3. **M. Bogyo**, G. von Degenfeld and G. Blum 'Imaging of Protease Activity in Live Cells Using Activity Based Probes' US 8,968,700 B2. Mar 3, 2015

4. **M. Bogyo** and A. Berger "Selective inhibitors and active site probes of caspases" Utility filing US Patent 12/306,215.
5. **M. Bogyo**, M. Fonovic and S. Verhelst "Mild Chemically Cleavable Linker for Proteomic Applications" Utility filing 12/376,053.
6. **M. Bogyo**, G. Blum, A.B. Berger, Z. Chen, S.S. Gambhir "Probes for in vivo targeting of active cysteine proteases" US 8,343,458 B2 Jan 1, 2013.
7. **M. Bogyo**, K.C. Garcia, P. Lupardus, A. Shen "Inducible self-cleaving protease tag and method of purifying recombinant proteins using the same" WO 2011/034622 A3 Mar 24, 2011.
8. **M. Bogyo**, J. Lee, Yin F., L.E. Edgington. "Specific inhibitors and active site probes for legumain" WO 2011/075678 A1. June 23, 2011.
9. **M. Bogyo**, A.W. Puri, A. Shen "Small molecule inhibitors of Clostridium difficile toxin" US Provisional Application 61/415,669.
10. **Bogyo**, M. Verdoes, L.E. Edgington "Non-peptidic quenched fluorescent imaging probes" WO 2012/118715 A2 Sept. 7, 2012.
11. **M. Bogyo**, M. Verdoes "Optical Probes for imaging of tumor margins and sites of inflammation" US Patent 14/777,024, International Filings
12. **M. Bogyo**, M. Verdoes, L. Ofori, N. Withana. "Protease-activated contrast agents for in vivo imaging" Patent US 2024/0025945 A1
13. **M. Bogyo**, A. Puri, T. Oresic Bender "Non-antibiotic drugs for the treatment of Clostridium difficile infection" PCT/US2015/037242
14. Phelps, D., Johnson, G., Bartik, M.M., Bensen, E., Halverson, D.J., and **Bogyo, M.** "Cysteine protease inhibitors" WO 2017/053864 A1.
15. Shabat, D., Green, O., **Bogyo, M.** "Chemiluminescent probes for Tuberculosis" US 2023/0094870 A1. Published Mar 30, 2023.
16. **M. Bogyo**, J. Widen, J. Yim "Multi-enzyme-activated imaging probes and related compositions and methods" WO 2020/081454 A1. April 23, 2020.

PUBLICATIONS

1. Wenzel, T.J., **Bogyo, M.**, and Lebeau, E., (1994) Lanthanide-cyclodextrin complexes as probes for elucidating optical purity by NMR spectroscopy. *Journal of the American Chemical Society*, 116, 4858-4865.
2. Wiertz, E. J. H. J., Jones, T. R., Sun, L., **Bogyo, M.**, Geuze, H. J., and Ploegh, H. L. (1996) The human cytomegalovirus US11 gene product dislocates MHC class I heavy chains from the endoplasmic reticulum to the cytosol. *Cell*, 84, 769-779.
3. Wiertz, E. J. H. J., Tortorella, D., **Bogyo, M.**, Yu, J., Mothes, W., Jones, T. R., Rapoport, T. A., and Ploegh, H. L. (1996) Sec61-mediated transfer of a membrane protein from the endoplasmic reticulum to the proteasome for destruction. *Nature*, 384, 432-438.
4. Jallepalli, P. and **Bogyo, M.** (1997) A degrading Business: the biology of proteolysis. *Trends in Cell Biology*, 7, 333-335.
5. **Bogyo, M.**, McMaster, J. S., Gaczynska, M., Tortorella, D., A.L. Goldberg and Ploegh, H. L. (1997) Covalent modification of the active site threonine of proteasomal β -subunits and the *Escherichia coli* homologue HslV by a new class of inhibitors. *Proceedings of the National Academy of Sciences USA*, **94**, 6629-6634.
6. **Bogyo, M.**, Gaczynska, M., Ploegh, H. L. (1997) Proteasome inhibitors and antigen presentation. *Biopolymers Peptide Science*. 47, 269-280.
7. Ruepp, A., Eckerskorn, C., **Bogyo, M.**, and Baumeister, W. (1998) Proteasome function is dispensable under normal but not under heat shock conditions in *Thermoplasma acidophilum*. *FEBS Lett.*, 425, 87-90.
8. Glas, R., **Bogyo, M.**, McMaster, J. S., Gaczynska, M., Ploegh, H. L. (1998) A proteolytic system that compensates for loss of proteasome function. *Nature*, 392, 618-622.
9. **Bogyo, M.**, McMaster, J. S., Shin, S., Ploegh, H. L. (1998) Substrate binding and sequence selectivity of the proteasome revealed by active site directed affinity probes. *Chemistry and Biology*, **5**, 307-320.

10. **Bogyo, M.** and Ploegh, H. L. (1998) A protease draws first blood. *Nature*. 396, 625-27.
11. Selzer, P. M., Pingel, S., Hsieh, I., Ugele, B., Chan, V. J., Engel, J. C., **Bogyo, M.**, Russell, D. G., Sakanari, J. A., and McKerrow, J. H. (1999) Cysteine protease inhibitors as chemotherapy: Lessons from a parasite target. *Proceedings of the National Academy of Sciences, USA*, 96, 11015-11022.
12. Schmidtke, G., Holzhütter, **Bogyo, M.**, Kairies, N., Groll, M., de Giuli, R., Emch, S., and Groettrup, M. (1999) How an inhibitor of the HIV-1 protease modulates proteasome activity. *J. Biol. Chem.*, 274. 35734-35740.
13. **Bogyo, M.**, Verhelst, S., Bellingard-Dubouchaud, V., Tobe, S., and Greenbaum, D. (2000). Selective targeting of lysosomal cysteine proteases with radio-labeled substrate analogs. *Chemistry and Biology*. 7, 27-38.
14. Caffrey, C.R., Mathieu, M.A., Gaffney, A.M., Salter, J.P., Sajid, M., Lucas, K.D., Franklin, C., **Bogyo, M.**, and McKerrow, J.H. (2000) Identification of a cDNA encoding an active asparaginyl endopeptidase of *Schistosoma mansoni* and its expression in *Pichia pastoris*. *FEBS Letters*. 466. 1-5.
15. Greenbaum, D. Medzihradzky, K.F. Burlingame, A. and **Bogyo, M.** (2000) Epoxide Electrophiles as Activity-Dependent Cysteine Protease Profiling and Discovery Tools. *Chemistry and Biology*, 7, 569-581.
16. Wang, E., Kessler, B., Borodovsky, A., Cravatt, B., **Bogyo, M.**, Ploegh, H. L., and Glas, R., (2000) Integration of the ubiquitin-proteasome pathway with a cytosolic oligopeptidase activity. *Proceedings of the National Academy of Sciences, USA*, 97, 9990-9995.
17. Weihofen, A., Lemberg, M. K., Ploegh, H., **Bogyo, M.** and Martiglio, B. (2000) Release of signal peptide fragments into the cytosol requires intramembrane cleavage by a protease activity that is specifically blocked by a novel cysteine protease inhibitor. *J. Biol. Chem.*, 275, 30951-30956.
18. Li, J., Gao, J., Ortega, J., Nazif, T., Joss, L., **Bogyo, M.**, Steven, A., and Rechsteiner, M. (2001). Proteasome activation by 11S REG (PA28) homologs: lysine 188 substitutions convert the pattern of proteasome activation by REG γ to that of REGs α and β . *EMBO J.*, 20, 3359-3369.
19. Nazif, T., and **Bogyo, M.** (2001) Global analysis of proteasomal substrate specificity using positional-scanning libraries of covalent inhibitors. *Proceeding of the National Academy of Sciences, USA*, 98, 2967-2972.
20. Baruch, A., Greenbaum, D., Levy E.T., Nielsen P.A., Gilula, N.B., Kumar, N.M., and **Bogyo, M.** (2001) Defining a link between gap junction communication, proteolysis, and cataract formation. *J. Biol. Chem.*, 276, 28999-29006.
21. Caffrey, C.R., Hansel, E., Lucas, K.D., Brinen, L.S., Alvarez Hernandez, A., Cheng, J., Gwaltney, S.L.II, Roush, W.R., Stierhof, Y-D., **Bogyo, M.**, Steverding, D. and McKerrow, J.H. (2001) Active site mapping, biochemical properties and subcellular localization of rhodesain, the major cysteine protease of *Trypanosoma brucei* rhodesiense. *Molecular & Biochemical Parasitology*, 118, 61-73.
22. **Bogyo, M.**, and Wang, E.W.(2002) Proteasome Inhibitors: Complex tools for a Complex Enzyme. *Current Topics in Microbiology and Immunology*, 268, 185-208.
23. Mathieu, M.A., **Bogyo, M.**, Caffrey, C.R., Choe, Y., Lee, J., Chapman, H., Sajid, M., Craik, C.S., and McKerrow, J.H. (2002) Substrate specificity of schistosome versus human legumain determined by P1-P3 peptide libraries. *Molecular & Biochemical Parasitology*, 121, 99-105.
24. Greenbaum, D.C., Baruch, A., Hayrapetian, L., Medzihradzky, K.F., Darula, Z., Burlingame, A., and **Bogyo, M.** (2002) Chemical approaches for functionally probing the proteome. *Mol. Cell. Proteomics*, 1, 60-68.
25. Groll, M., Nazif, T., Huber, R., and **Bogyo, M.** (2002) Probing structural determinants distal to the site of hydrolysis that control substrate specificity of the 20S proteasome. *Chemistry and Biology*, 9, 655-662.
26. Greenbaum, D., Arnold, W., Lu, F., Hayrapetian, L., Baruch, A., Krumrine, J., Toba, S., Chehade, K., Bromme, D., Kuntz, I.D., and **Bogyo, M.** (2002) Small Molecule affinity fingerprinting: a tool for enzyme family sub-classification, target identification, and inhibitor design. *Chemistry and Biology*, 9, 1085-1094.

27. Greenbaum, D., Baruch, A., Grainger, M., Bozdech, Z., Medzihradzky, K., Engel, J., Holder, T., DeRisi, J., and **Bogyo, M.** (2002) A role for the cysteine protease falcipain 1 in host cell invasion by the malaria parasite, *Plasmodium falciparum*. *Science*, 298, 2002-2006.
28. Kessler, B., Hong, X., Petrovic, J., Borodovsky, A., Dantuma, N.P., **Bogyo, M.**, Overkleeft, H.S., Ploegh, H., and Glas, R. (2003) Pathways accessory to proteasomal proteolysis are less efficient in MHC class I antigen presentation. *J. Biol. Chem.* 278, 10013-21.
29. Mikolajczyk, J., Boatright, K.M., Stennicke, H.R., Nazif, T.M., Potempa, J., **Bogyo, M.**, and Salvesen, G.S. (2003) Sequential autocatalytic processing activates the zymogen of the caspase homolog Arg-gingipain. *J. Biol. Chem.* 278, 10458-64.
30. **Bogyo, M.** and Hurley, J. (2003) Proteomics and Genomics. *Current Opinions in Chemical Biology.* 7, 2-4.
31. Jeffery, D., and **Bogyo, M.** (2003) Chemical proteomics and applications to drug discovery. *Current Opinions in Biotechnology.* 14, 82-86.
32. Wang, C.C., Bozdech, Z., Liu, C., Harris, J., and **Bogyo, M.** (2003) Biochemical analysis of the *Tropanosoma brucei* proteasome. *J. Biol. Chem.* 278, 15800-8.
33. Rozman-Pungercar, J., Kopitar Jerala, N., **Bogyo, M.**, Turk, D., Vasiljeva, O., Stefe, I., Vandenabeele, P., Brömme, D., Puizdar, V., Fonovic, M., Trstenjak-Prebanda, M., Dolenc, I., Turk, V., and Turk, B. (2003) When reaction mechanism is more important than specificity: Inhibition of papain-like cysteine proteases and legumain by "caspase-specific" inhibitors. *Cell Death and Differentiation*, 10, 881-888.
34. Yasothornsrikul, S., Greenbaum, D., Medzihradzky, K., Toneff, T., Bunday, R., Miller, R., Schilling, B., Petermann, I., Dehnert, J., Logvinova, A., Goldsmith, P., Neveu, J., Lane, W., Gibson, G., Reinheckel, T., Peters, C., **Bogyo, M.***, and Hook, V. (2003) Cathepsin L in secretory vesicles functions as a prohormone-processing enzyme for production of the enkephalin peptide neurotransmitter. *Proceedings of the National Academy of Sciences, USA*, 100, 9590-9595. *Co-Senior Authors.
35. Sajid M., McKerrow J.H., Hansell E., Mathieu M.A., Lucas K.D., Hsieh I., Greenbaum D., **Bogyo M.**, Salter J.P., Lim K.C., Franklin C., Kim J.H., Caffrey C.R. (2003) Functional expression and characterization of *Schistosoma mansoni* cathepsin B and its trans-activation by an endogenous asparaginyl endopeptidase. *Mol. Biochem. Parasitol.*, 131, 65-75.
36. Li, Z., Yasuda, Y., Li, W., **Bogyo, M.**, Katz, N., Gordon, G., Fields, G.B., and Brömme, D. (2004) Regulation of collagenase activities of human cathepsins by glycosaminoglycans. *J Biol Chem.* 279, 5470-5479.
37. Baruch, A., Jeffery D., and **Bogyo, M.** (2004) Enzyme activity – it's all about image. *Trends in Cell Biology.* 14, 29-35.
38. Medzihradzky, K.F., Darula, Z., Perlson, E., Fainzilber, M., Chalkley, R.J., Ball, H., Greenbaum, D., **Bogyo, M.**, Tyson, D.R., Bradshaw, R.A., and Burlingame, A.L. (2004) O-Sulfonation of serine and threonine - mass spectrometric detection and characterization of a new posttranslational modification in diverse proteins throughout the eukaryotes. *Mol. Cell. Proteomics.* 3, 429-440.
39. Goulet, B., Baruch, A., Greenbaum, D., Moon, N.-S., Poirier, M., Erickson, A., **Bogyo, M.***, and Nepveu, A.* (2004) A role for the lysosomal cysteine protease cathepsin L in transcriptional regulation in the nucleus. *Molecular Cell.* 14, 207-219.
40. Joyce, J., Baruch, A., Chehade, K., Greenbaum, D., Meyer-Morse, N., Tsai, F.-Y., Greenbaum, D., Hager, J., **Bogyo, M.***, and Hanahan, D.* (2004) Cathepsin cysteine proteases are effectors of invasive growth and angiogenesis during multistage tumorigenesis. *Cancer Cell.* 5, 443-453. *Co-Senior Authors.
41. Yasuda Y, Li Z, Greenbaum D, Bogyo M, Weber E, Bromme D. (2004) Cathepsin V: A novel and potent elastolytic activity expressed in activated macrophages. *J Biol Chem.* 279, 36761-36770.

42. Eksi, S., Czesny, B., Greenbaum, D.C., **Bogyo, M.**, and Williamson, K. (2004) Targeted disruption of *Plasmodium falciparum* cysteine protease, falcipain 1, reduces oocyst production, not erythrocytic stage growth. *Molecular Microbiology*. 53, 243-250.
43. **Bogyo, M.**, Baruch A., Jeffery, D.A., Greenbaum, D., Ovaa, H., Borodovsky, A., and Kessler, B. (2004) Applications of chemical probes of proteolytic activity. *Current Protocols in Protein Science*. Sep;Chapter 21:Unit 21.17
44. Van Der Hoorn RA, Leeuwenburgh MA, **Bogyo M**, Joosten MH, Peck SC. (2004) Activity profiling of papain-like cysteine proteases in plants. *Plant Physiol*. 135, 1170-1178.
45. Oleksy A, Golonka E, Banbula A, Szmyd G, Moon J, Kubica M, Greenbaum D, **Bogyo M**, Foster TJ, Travis J, Potempa J. (2004) Growth phase-dependent production of a cell wall-associated elastinolytic cysteine proteinase by *Staphylococcus epidermidis*. *Biol Chem*. 385, 525-535.
46. Hook V, Yasothornsrikul S, Greenbaum D, Medzihradzky KF, Troutner K, Toneff T, Bunday R, Logrinova A, Reinheckel T, Peters C, **Bogyo M**. (2004) Cathepsin L and Arg/Lys aminopeptidase: a distinct prohormone processing pathway for the biosynthesis of peptide neurotransmitters and hormones. *Biol Chem*. 385, 473-480.
47. Snipas SJ, Wildfang E, Nazif T, Christensen L, Boatright KM, **Bogyo M**, Stennicke HR, Salvesen GS. (2004) Characteristics of the caspase-like catalytic domain of human paracaspase. *Biol Chem*. 385, 1093-1098.
48. Berger, A.B., Vitorino, P.M., and **Bogyo, M.** (2004) Activity-based protein profiling: applications to biomarker discovery, *in vivo* imaging and drug discovery. *Am. J. of Pharmacogenomics*, 4, 371-381.
49. Borodovsky A, Ovaa H, Meester WJ, Venanzi ES, **Bogyo M**, Hekking BG, Ploegh HL, Kessler BM, Overkleeft HS. (2005) Small-molecule inhibitors and probes for ubiquitin- and ubiquitin-like-specific proteases. *ChemBioChem*, 6, 287-291.
50. Chehade, K.A.H., Baruch, A., Verhelst, S.H.L., and **Bogyo, M.** (2005) An improved preparation of the activity-based probe JPM-OEt and *in situ* applications. *Synthesis*, 2, 240-244.
51. Verhelst, S.H.L., and **Bogyo, M.** (2005) Chemical proteomics applied to target identification and drug discovery. *BioTechniques*, 38, 175-177.
52. Verhelst, S.H.L., and **Bogyo, M.** (2005) Solid Phase Synthesis of double headed epoxysuccinyl activity based probes for selective targeting of papain family cysteine proteases. *ChemBioChem*, 6, 824-827.
53. Phillips, C.I., and **Bogyo, M.** (2005) Proteomics meets microbiology: technical advances in the global mapping of protein expression and function. *Cellular Microbiology* 7(8) 1061-76.
54. Kato, D., Boatright, K.M., Berger, A.B., Nazif, T., Blum, G., Ryan, C., Chehade, K.A.H., Salvesen, G., and **Bogyo, M.** (2005) Activity based probes that target diverse cysteine protease families. *Nature Chemical Biology*. 1, 33-38.
55. Blum, G., Mullins, S.R., Keren, K., Fonovic, J., Jedeszko, C., Rice, M.J., Sloane, B.F., and **Bogyo, M.** (2005) Dynamic imaging of protease activity with fluorescently quenched activity-based probes. *Nature Chemical Biology*, 1, 203-209.
56. Hook V, Toneff T, **Bogyo M**, Greenbaum D, Medzihradzky KF, Neveu J, Lane W, Hook G, and Reisine T. (2005) Inhibition of cathepsin B reduces beta-amyloid production in regulated secretory vesicles of neuronal chromaffin cells: evidence for cathepsin B as a candidate beta-secretase of Alzheimer's disease. *Biol. Chem*. 386, 931-940.
57. Kato, D., Verhelst, S.H.L., Sexton, K.B. and **Bogyo, M.** (2005) A General Solid Phase Method for the Preparation of Diverse Azapeptide Probes Directed Against Cysteine Proteases. *Organic Letters*, 7, 5649-5652.

58. **Bogyo M.** (2005) Screening for selective small molecule inhibitors of the proteasome using activity-based probes. *Methods Enzymol.* 399, 609-22.
59. Choe Y., Leonetti F., Greenbaum D.C., Lecaille F., **Bogyo M.**, Bromme D., Ellman J.A., Craik C.S. (2006) Substrate profiling of cysteine proteases using a combinatorial peptide library identifies functionally unique specificities. *J Biol Chem.* 281, 12824-12832.
60. Verhelst, S.H.L., Witte, M.D., Arastu-Kapur, S., Fonovic M., and **Bogyo, M.** (2006) Novel aza peptide inhibitors and active site probes of papain family cysteine proteases. *ChemBioChem*, 7, 943-950.
61. Yuan, F., Verhelst, S., Blum, G., Coussens, L., and **Bogyo, M.** (2006) A selective activity-based probe for the papain family cysteine protease dipeptidyl peptidase I/cathepsin C. *J. Am. Chem. Soc.* 128, 5616-5617.
62. Pan Z, Jeffery DA, Chehade K, Beltman J, Clark JM, Grothaus P, **Bogyo M**, Baruch A. (2006) Development of activity-based probes for trypsin-family serine proteases. *Bioorg. Med. Chem. Lett.* 16, 2882-2885.
63. **Bogyo M.** (2006) Metallo proteases see the light. *Nature Chem. Bio.* 2, 229-230.
64. Vasiljeva O, Papazoglou A, Kruger A, Brodoefel H, Korovin M, Deussing J, Augustin N, Nielsen BS, Almholt K, **Bogyo M**, Peters C, Reinheckel T. (2006) Tumor cell-derived and macrophage-derived cathepsin B promotes progression and lung metastasis of mammary cancer. *Cancer Research*, 66, 5242-5250.
65. Berger A.B., Witte M., Sadaghiani A.M., Sexton K.M.B., Denault J.B., Salvesen G.S., and **Bogyo, M.** (2006) Identification of early intermediates of caspase activation during intrinsic apoptosis using selective inhibitors and activity based probes. *Molecular Cell*, 23, 509-521.
66. Denault, J.B., Bekes M., Sexton K.M.B., Scott F.L., **Bogyo M.**, and Salvesen G.S. (2006) Conformational mobility in the activation of executioner caspases. *Molecular Cell*, 23, 523-533.
67. Pandey KC, Singh N, Arastu-Kapur S, Bogyo M, Rosenthal PJ. Falstatin, a cysteine protease inhibitor of Plasmodium falciparum, facilitates erythrocyte invasion. (2006) *PLoS Pathog.* Nov;2(11):e117.
68. Sadaghiani, M., Verhelst, S.H.L. and **Bogyo, M.** (2006) Solid phase methods for the preparation of epoxysuccinate-based inhibitors of cysteine proteases. *J. Comb. Chem.* 8, 802-804.
69. Berger, A.B., Sexton, K.B., and **Bogyo, M.** (2006) Commonly used caspase inhibitors designed based on substrate specificity profiles lack selectivity. *Cell Research*, 16, 961-963.
70. Sexton, K.B., Kato, D., Berger, A.B., Fonovic, M., Verhelst, S.H.L., and **Bogyo, M.** (2007) Specificity of aza-peptide electrophile activity based probes of caspases. *Cell Death and Differentiation.* 14, 727-732.
71. Sexton, K.B., Witte, M.D., Blum, G., and **Bogyo, M.** (2007) Design of cell permeable, fluorescent activity-based probes of the lysosomal cysteine protease asparaginyl endopeptidase (AEP)/legumain. *Bioorg. Med. Chem.* 17, 649-653.
72. Sadaghiani, A.M., Verhelst, S.H.L., and **Bogyo, M.** (2007) Tagging and detection strategies for activity-based proteomics. *Cur. Opin. Chem. Bio.* 11, 20-28.
73. Cuerrier, D., Moldoveanu, T., Campbell, R.L., Kelly, J., Yoruk, B., Verhelst, S.H., Greenbaum, D., **Bogyo, M.**, Davies, P.L. (2007) Development of calpain-specific inactivators by screening of positional-scanning epoxide libraries. *J Biol Chem.* 282, 9600- 9611.
74. Verhelst, S.H.L., Fonovic, M. and **Bogyo, M.** (2007) A mild Chemically Cleavable linker for proteomic applications *Agnew. Chem.* 46, 1284-1286.
75. Fonovic, M., and **Bogyo, M.** (2007) Activity based probes for proteases: applications Applications to Biomarker Discovery, Molecular Imaging and Drug Screening. *Cur, Pharm. Des.* 13, 253-261.

76. Teo CF, Zhou XW, **Bogyo, M**, Carruthers VB. (2007) Cysteine protease inhibitors block *Toxoplasma gondii* microneme secretion and cell invasion. *Antimicrob Agents Chemother.* 51, 679-88.
77. **Bogyo, M** and Cravatt BF. (2007) Genomics and proteomics From genes to function: advances in applications of chemical and systems biology. *Curr Opin Chem Biol.* 11, 1-3.
78. Sojka D, Hajdusek O, Dvorak J, Sajid M, Franta Z, Schneider EL, Craik CS, Vancova M, Buresova V, **Bogyo M**, Sexton KB, McKerrow JH, Caffrey CR, Kopacek P. (2007) IrAE - An asparaginyl endopeptidase (legumain) in the gut of the hard tick *Ixodes ricinus*. *Int J Parasitol.* 37, 713-724.
79. Sadaghiani, A.M., Verhelst, S.H.L., Gocheva, V., Hill, K., Majerova, E., Stinson, S. Joyce, J.A. and **Bogyo, M.** (2007) Design, Synthesis and Evaluation of *in vivo* Potency and Selectivity of Epoxysuccinyl-Based Inhibitors of Papain Family Cysteine Proteases. *Chem. Biol.* 14, 499-511.
80. Rose PP, **Bogyo M**, Moses AV, Fruh K. (2007) Insulin-like Growth Factor II Receptor-mediated intracellular Retention of Cathepsin B is essential for transformation of endothelial cells by Kaposi's sarcoma associated herpesvirus. *J. Virol.* 81, 8050-8062.
81. Burster T, Giffon T, Dahl ME, Bjorck P, **Bogyo M**, Weber E, Mahmood K, Lewis DB, Mellins ED. (2007) Influenza A virus elevates active cathepsin B in primary murine DC. *Int Immunol.* 19, 645-55.
82. Bell-McGuinn, KM, Garfall, AL, **Bogyo, M**, Hanahan, D. and Joyce, JA. (2007) Inhibition of cysteine cathepsin protease activity enhances chemotherapy regimens by decreasing tumor growth and invasiveness in a mouse model of multistage cancer. *Cancer Res.* 67, 7378-7385.
83. Fonović, M., Verhelst, S.H.L., Sorum, M.T., and **Bogyo, M.** (2007) Proteomic evaluation of chemically cleavable activity based probes. *Mol. Cell. Proteomics* 6, 1761-1770.
84. Blum, G., von Degenfeld, G., Merchant, M.J., Blau, H.M., and **Bogyo, M.** (2007) Optical Imaging of Cysteine Protease Activity in Living Subjects Using Quenched Near Infrared Fluorescent Activity Based Probes (NIRF-qABPs). *Nat. Chem. Biol.* 3, 668-677.
85. Goulet B, Sansregret L, Leduy L, **Bogyo M**, Weber E, Chauhan SS, Nepveu A. (2007) Increased expression and activity of nuclear cathepsin L in cancer cells suggests a novel mechanism of cell transformation. *Mol Cancer Res.* 5, 899-907.
86. Ponder EL, **Bogyo M.** (2007) Ubiquitin-like modifiers and their deconjugating enzymes in medically important parasitic protozoa. *Eukaryot Cell.* 6, 1943-52.
87. Arastu-Kapur S., Ponder E.L., Fonovic U., Yeoh S., Yuan F., Fonovic, M., Grainger M., Phillips C.I., Powers J.C., and **Bogyo M.** (2008) A small molecule screen identifies proteases that regulate erythrocyte rupture by the human malaria parasite *Plasmodium falciparum* *Nature Chemical Biology*, 4, 203-213.
88. Paulick MG, **Bogyo M.** (2008) Application of activity-based probes to the study of enzymes involved in cancer progression. *Curr Opin Genet Dev.* 18, 97-106.
89. Yang Z, Fonović M, Verhelst SH, Blum G, **Bogyo M.** (2009) Evaluation of alpha,beta-unsaturated ketone-based probes for papain-family cysteine proteases. *Bioorg Med Chem.* 17, 1071-8
90. Schurigt U, Sevenich L, Vannier C, Gajda M, Schwinde A, Werner F, Stahl A, von Elverfeldt D, Becker AK, **Bogyo M**, Peters C, Reinheckel T. (2008) Trial of the cysteine cathepsin inhibitor JPM-OEt on early and advanced mammary cancer stages in the MMTV-PyMT-transgenic mouse model. *Biol Chem.* 389, 8, 1067-74.
91. Obermajer N, Svajger U, **Bogyo M**, Jeras M, Kos J. (2008) Maturation of dendritic cells depends on proteolytic cleavage by cathepsin X. *J Leukoc Biol.* 84, 1306-1315.
92. Jevnikar Z, Obermajer N, **Bogyo M**, Kos J. (2008) The role of cathepsin X in the migration and invasiveness of T lymphocytes. *J Cell Sci.* 121, 2652-61.

93. Lupardus PJ, Shen A, **Bogyo M**, Garcia KC. (2008) Small molecule-induced allosteric activation of the *Vibrio cholerae* RTX cysteine protease domain. *Science*. 322, 265-8.
94. Shen A, **Bogyo M**. (2008) Friend or foe? Turning a host defense protein into a pathogen's accomplice. *Chem Biol*. 15, 879-80.
95. Fonović M, **Bogyo M**. (2008) Activity-based probes as a tool for functional proteomic analysis of proteases. *Expert Rev Proteomics*. 5, 721-30.
96. Kaschani F, Verhelst SH, van Swieten PF, Verdoes M, Wong CS, Wang Z, Kaiser M, Overkleeft HS, **Bogyo M**, van der Hoorn RA. (2008) Minitags for small molecules: detecting targets of reactive small molecules in living plant tissues using 'click-chemistry' *Plant J*. 57, 373-385.
97. Pungercar JR, Caglic D, Sajid M, Dolinar M, Vasiljeva O, Pozgan U, Turk D, Bogyo M, Turk V, Turk B. (2009) Autocatalytic processing of procathepsin B is triggered by proenzyme activity. *FEBS J*. 276, 660-8.
98. Liz MA, Fleming CE, Nunes AF, Almeida MR, Mar FM, Choe Y, Craik CS, Powers JC, **Bogyo M**, Sousa MM. (2009) Substrate specificity of transthyretin: identification of natural substrates in the nervous system. *Biochem J*. 419, 467-474.
99. Edgington L, Berger AB, Blum G, Albrow VE, Lineberry N, **Bogyo M**. (2009) Imaging apoptosis *in vivo* using caspase-targeted activity based probes, *Nature Medicine*, 15, 967-973
100. Shen A, Lupardus PJ, Albrow VE, Guzzetta A, Powers JC, Garcia KC, **Bogyo M**. (2009) Mechanistic and structural insights into the proteolytic activation of *Vibrio cholerae* MARTX toxin. *Nat Chem Bio*, 24, 469-478.
101. Obermajer N, Jevnikar Z, Doljak B, Sadaghiani AM, **Bogyo M**, Kos J (2009) Cathepsin X-mediated beta2 integrin activation results in nanotube outgrowth. *Cell Mol Life Sci*. 66, 1126-34.
102. Cavallo-Medved D, Rudy D, Blum G, **Bogyo M**, Caglic D, Sloane BF. (2009) Live-cell imaging demonstrates extracellular matrix degradation in association with active cathepsin B in caveolae of endothelial cells during tube formation. *Exp Cell Res*. 315,1234-46.
103. Barbero S, Mielgo A, Torres V, Teitz T, Shields DJ, Mikolon D, **Bogyo M**, Barilà D, Lahti JM, Schlaepfer D, Stupack DG. (2009) Caspase-8 association with the focal adhesion complex promotes tumor cell migration and metastasis. *Cancer Res*. 69, 3755-63.
104. Beckham SA, Piedrafita D, Phillips CI, Samarawickrema N, Law RH, Smooker PM, Quinsey NS, Irving JA, Greenwood D, Verhelst SH, **Bogyo M**, Turk B, Coetzer TH, Wijeyewickrema LC, Spithill TW, Pike RN. (2009) A major cathepsin B protease from the liver fluke *Fasciola hepatica* has atypical active site features and a potential role in the digestive tract of newly excysted juvenile parasites. *Int J Biochem Cell Biol*. 41, 1601-12.
105. Chang SH, Kanasaki K, Gocheva V, Blum G, Harper J, Moses MA, Shih SC, Nagy JA, Joyce J, **Bogyo M**, Kalluri R, Dvorak HF. (2009) VEGF-A induces angiogenesis by perturbing the cathepsin-cysteine protease inhibitor balance in venules, causing basement membrane degradation and mother vessel formation. *Cancer Res*. 69, 4537-44.
106. Larson ET, Parussini F, Huynh MH, Giebel JD, Kelley AM, Zhang L, **Bogyo M**, Merritt EA, Carruthers VB. (2009) *Toxoplasma gondii* cathepsin I is the primary target of the invasion inhibitory compound LHSV. *J Biol Chem*. 284, 26839-50.
107. Puri AW, **Bogyo M**. (2009) Using Small Molecules To Dissect Mechanisms of Microbial Pathogenesis. *ACS Chem Biol*. 4, 603-16.
108. Lee JT, Chen DY, Yang Z, Ramos AD, Hsieh JJ, **Bogyo M**. (2009) Design, syntheses, and evaluation of Taspase1 inhibitors. *Bioorg Med Chem Lett*. 19, 5086-90.

109. Blum G, Weimer RM, Edgington LE, Adams W, **Bogyo M.** (2009) Comparative assessment of substrates and activity based probes as tools for non-invasive optical imaging of cysteine protease activity. *PLoS One* 4(7):e6374.
110. Zhang J, Fonovic M, Suyama K, **Bogyo M,** Scott MP. Rab35 controls actin bundling by recruiting fascin as an effector protein. (2009) *Science* 325, 1250-4.
111. Horn M, Nussbaumerová M, Sanda M, Kovárová Z, Srba J, Franta Z, Sojka D, **Bogyo M,** Caffrey CR, Kopáček P, Mares M. Hemoglobin digestion in blood-feeding ticks: mapping a multipeptidase pathway by functional proteomics. (2009) *Chem. Biol.* 16, 1053-63.
112. Shen A., Lupardus, P.J., Morell, M., Ponder, E.L., Sadaghiani, M., Garcia, K.C., **Bogyo, M.** (2009) Simplified, enhanced protein purification using an inducible, autoprocessing enzyme tag. *PLoS One* 4(12):e8119.
113. Ravindran S, Lodoen MB, Verhelst SH, **Bogyo M,** Boothroyd JC. (2009) 4-bromophenacyl bromide specifically inhibits rhoptry secretion during *Toxoplasma* invasion. *PLoS One.* 4(12):e8143.
114. Drag M, **Bogyo M,** Ellman JA, Salvesen GS. (2010) Aminopeptidase fingerprints, an integrated approach for identification of good substrates and optimal inhibitors. *J Biol Chem.* 2010 Jan 29;285(5):3310-8.
115. **Bogyo M.** (2010) Finding enzymes that are actively involved in cancer. *Proc Natl Acad Sci U S A.* 2010 Feb 9;107(6):2379-80.
116. Lee, J, and **Bogyo, M.** (2010) Development of near-infrared fluorophore (NIRF)-labeled activity-based probes for *in vivo* imaging of legumain. *ACS Chem. Bio.* 5(2):233-43.
117. Tedelind S, Poliakova K, Valeta A, Hunegnaw R, Yemanaberhan EL, Heldin NE, Kurebayashi J, Weber E, Kopitar-Jerala N, Turk B, **Bogyo M,** Brix K. (2010) Nuclear cysteine cathepsin variants in thyroid carcinoma cells. *Biol Chem.* 391(8):923-35.
118. Staudt ND, Aicher WK, Kalbacher H, Stevanovic S, Carmona AK, **Bogyo M,** Klein G. (2010) Cathepsin X is secreted by human osteoblasts, digests CXCL-12 and impairs adhesion of hematopoietic stem and progenitor cells to osteoblasts. *Haematologica.* 95, 1452-60.
119. Konjar S, Yin F, **Bogyo M,** Turk B, Kopitar-Jerala N. (2010) Increased nucleolar localization of SpiA3G in classically but not alternatively activated macrophages. *FEBS Lett.* 584(11):2201-6.
120. Deu E, Yang Z, Wang F, Klemba M, **Bogyo M.** (2010) Use of activity-based probes to develop high throughput screening assays that can be performed in complex cell extracts. *PLoS One.* 5(8). pii: e11985.
121. Deu E, Leyva M, Albrow VE, Rice MJ, Ellman JA, **Bogyo M.** (2010) Functional studies of *Plasmodium falciparum* dipeptidyl aminopeptidase I (DPAP1) using small molecule inhibitors and active site probe. *Chemistry and Biology.* 17, 808-19.
122. Wang F, Krai P, Deu E, Bibb B, Lauritzen C, Pedersen J, **Bogyo M,** Klemba M. (2010) Biochemical characterization of *Plasmodium falciparum* dipeptidyl aminopeptidase 1. *Mol Biochem Parasitol.* 175(1):10-20.
123. Puri AW, Lupardus, PJ, Deu E, Albrow VA, Garcia KC, **Bogyo M.*** and Shen A. (2010) Rational Design of Inhibitors and Activity-Based Probes Targeting *Clostridium difficile* Virulence Factor TcdB. *Chemistry and Biology* 17, 1201-2011. *Senior Author
124. Bowyer PW, Simon GM, Cravatt BF, **Bogyo M.** (2011) Global profiling of proteolysis during rupture of *P. falciparum* from the host erythrocyte. *Mol Cell Proteomics.* 10, M110.001636
125. Shen A., Lupardus, P.J., Puri, A.W., Albrow, V.E., Gersch, M.M., Garcia, K.C., and **Bogyo, M.** (2011) Defining an allosteric circuit in the cysteine protease domain of *Clostridium difficile* glucosylating toxins. *Nature Structure and Molecular Biology.* 6, 415-419.

126. Paulick, M and **Bogyo, M.** (2011) Development of Activity-Based Probes for Cathepsin X. *ACS Chem Biol.* 6, 563-572.
127. Mahajan SS, Deu E, Lauterwasser EM, Leyva MJ, Ellman JA, **Bogyo M***, Renslo AR. (2011) A fragmenting hybrid approach for targeted delivery of multiple therapeutic agents to the malaria parasite. *ChemMedChem.* 6, 415-419. *Co-senior Author.
128. Ewald SE, Engel A, Lee J, Wang M, **Bogyo M**, Barton GM. (2011) Nucleic acid recognition by Toll-like receptors is coupled to stepwise processing by cathepsins and asparagine endopeptidase. *J Exp Med.* 208, 643-51.
129. Boutté AM, Friedman DB, **Bogyo M**, Min Y, Yang L, Lin PC. (2011) Identification of a myeloid-derived suppressor cell cystatin-like protein that inhibits metastasis. *FASEB J.* 25, 2626-2637.
130. Ponder EL, Albrow VE, Leader BA, Békés M, Mikolajczyk j, Pečar Fonović U, Shen A, Drag M, Xiao J, Deu E, Campbell AJ, Powers JC, Salvesen GS, and **Bogyo M** (2011) Functional characterization of a SUMO deconjugating protease of *Plasmodium falciparum* using newly identified small molecule inhibitors. *Chemistry and Biology*, 18, 711-21.
131. Albrow VE, Ponder EL, Fasci D, Békés M, Deu E, Salvesen GS, and **Bogyo M.** (2011) Development of small molecule inhibitors and probes of human SUMO deconjugating proteases (SENPs). *Chemistry and Biology*, 18, 722-32.
132. Hall CI, Reese M, Weerapana E, Child MA, Bowyer PW, Albrow VE, Haraldsen JG, Phillips MR, Deu E, Ward GE, Cravatt BF, Boothroyd JC, and **Bogyo M.** (2011) A chemical genetic screen identifies Toxoplasma DJ-1 as a regulator of parasite secretion and invasion. *Proc. Natl. Acad. Sci. USA* 108, 10568-73.
133. Li H, Child MA, **Bogyo M.** (2011) Proteases as regulators of pathogenesis: Examples from the apicomplexa. *Biochim Biophys Acta.* 1824, 177-85.
134. Cattaruzza F, Lyo V, Jones E, Pham D, Hawkins J, Kirkwood K, Valdez-Morales E, Ibeakanma C, Vanner SJ, **Bogyo M**, Bunnett NW. (2011). Cathepsin S Is Activated During Colitis and Causes Visceral Hyperalgesia by a PAR(2)-Dependent Mechanism in Mice. *Gastroenterology.* 141, 1864-1874.
135. Kalińska M, Kantyka T, Greenbaum DC, Larsen KS, Władyka B, Jabaiah A, **Bogyo M**, Daugherty PS, Wysocka M, Jaros M, Lesner A, Rolka K, Schaschke N, Stennicke H, Dubin A, Potempa J, Dubin G. (2011) Substrate specificity of Staphylococcus aureus cysteine proteases - Staphopains A, B and C. *Biochimie.* 94, 318-27.
136. Mikhaylov G, Mikac U, Magaeva AA, Itin VI, Naiden EP, Psakhye I, Babes L, Reinheckel T, Peters C, Zeiser R, **Bogyo M**, Turk V, Psakhye SG, Turk B, Vasiljeva O. (2011) Ferri-liposomes as an MRI-visible drug-delivery system for targeting tumours and their microenvironment. *Nature Nanotechnology.* 6, 594-602.
137. Tedelind S, Jordans S, Resemann H, Blum G, **Bogyo M**, Führer D, Brix K. (2011) Cathepsin B trafficking in thyroid carcinoma cells. *Thyroid Res.* Aug 3;4 Suppl 1:S2.
138. Li J, Hsu HC, Yang P, Wu Q, Li H, Edgington LE, **Bogyo M**, Kimberly RP, Mountz JD. (2011) Treatment of arthritis by macrophage depletion and immunomodulation: Testing an apoptosis-mediated therapy in a humanized death receptor mouse model. *Arthritis Rheum.* 64, 1098-109.
139. Ren G, Blum G, Liu H, Gheysens O, Miao Z, Gambhir S, **Bogyo M**, and Cheng Z. (2011) Non-invasive imaging of cysteine cathepsin activity in solid tumors using a ⁶⁴Cu-labeled activity-based probe. *PLoS One.* 6(11):e28029.
140. Edgington LE, Verdoes M, **Bogyo M.** (2011) Functional imaging of proteases: recent advances in the design and application of substrate and activity-based probes. *Current Opinions in Chemical Biology.* 15, 798-805.
141. Deu E, Verdoes M, **Bogyo M.** (2012) New tools for dissecting protease function: implications for inhibitor design, drug discovery and probe development. *Nature Structure and Molecular Biology.* 19, 9-16.

142. Lee J and **Bogyo M**. (2012) Synthesis and evaluation of aza-peptidyl inhibitors of the lysosomal asparaginyl endopeptidase, legumain. *Bioorg. Med. Chem. Lett.* 22, 1340-3.
143. Ferreira KS, Kreutz C, Macnelly S, Neubert K, Haber A, **Bogyo M**, Timmer J, Borner C. (2012) Caspase-3 feeds back on caspase-8, Bid and XIAP in type I Fas signaling in primary mouse hepatocytes. *Apoptosis.* 17,503-15.
144. Withana NP, Blum G, Sameni M, Slaney C, Anbalagan A, Olive MB, Bidwell BN, Edgington L, Wang L, Moin K, Sloane BF, Anderson RL, **Bogyo MS**, Parker BS. (2012) Cathepsin B inhibition limits bone metastasis in breast cancer. *Cancer Res.* 72, 1199-209.
145. Fritz HM, Bowyer PW, **Bogyo M**, Conrad PA, Boothroyd JC. Proteomic analysis of fractionated toxoplasma oocysts reveals clues to their environmental resistance. (2012) *PLoS One.* 2012;7(1):e29955.
146. Edgington LE, van Raam BJ, Verdoes M, Wierschem C, Salvesen GS, and **Bogyo M** (2012) An optimized activity-based probe for the study of caspase-6 activation. *Chemistry & Biology.* 19, 340-52.
147. Richau KH, Kaschani F, Verdoes M, Pansuriya TC, Niessen S, Stüber K, Colby T, Overkleeft HS, **Bogyo M**, Van der Hoorn RA. (2012) Subclassification and biochemical analysis of plant papain-like cysteine proteases displays subfamily-specific characteristics. *Plant Physiol.* 158, 1583-99.
148. Cutter JL, Cohen NT, Wang J, Sloan AE, Cohen AR, Panneerselvam A, Schluchter M, Blum G, **Bogyo M**, Basilion JP. (2012) Topical Application of Activity-based Probes for Visualization of Brain Tumor Tissue. *PLoS One.* 7(3):e33060.
149. Verdoes M, Edgington LE, Scheeren F, Leyva M, Blum G, Bachman MH, Ellman JA, and **Bogyo M** (2012) A non-peptidic cathepsin S activity-based probe for non-invasive optical imaging of tumor associated macrophages. *Chemistry and Biology.* 9, 619-28.
150. Puri AW, Broz P, Shen, A, Monack DM, **Bogyo M**. (2012) An activity-based probe reveals caspase-1 activity is required to bypass apoptosis upon bacterial infection. *Nature Chemical Biology* 8, 745-7.
151. Goussetis DJ, Gounaris E, Wu EJ, Vakana E, Sharma B, **Bogyo M**, Altman JK, Plataniias LC. (2012) Autophagic degradation of the BCR-ABL oncoprotein and generation of antileukemic responses by arsenic trioxide. *Blood.* 120, 3555-62.
152. Lyo V, Cattaruzza F, Kim TN, Walker AW, Paulick M, Cox D, Cloyd J, Buxbaum J, Ostroff J, **Bogyo M**, Grady EF, Bunnett NW, Kirkwood KS. (2012) Active Cathepsins B, L and S in Murine and Human Pancreatitis. *Am J Physiol Gastrointest Liver Physiol.* 303, G894-903.
153. Salvesen GS, **Bogyo M**. (2012) Highlight: The universe of proteolytic networks and mechanisms. *Biol Chem.* 393, 841.
154. Sztukowska M, Veillard F, Potempa B, **Bogyo M**, Enghild JJ, Thogersen IB, Nguyen KA, Potempa J. (2012) Disruption of gingipain oligomerization into non-covalent cell-surface attached complexes. *Biol Chem.* 393, 971-7.
155. Misas-Villamil JC, Toenges G, Kolodziejek I, Sadaghiani AM, Kaschani F, Colby T, **Bogyo M**, van der Hoorn RA. (2012) Activity profiling of vacuolar processing enzymes reveals a role for VPE during oomycete infection. *Plant J.* 73, 689-700.
156. Stolze SC, Deu E, Kaschani F, Li N, Florea BI, Richau KH, Colby T, van der Hoorn RA, Overkleeft HS, **Bogyo M**, Kaiser M. (2012) The antimalarial natural product symprostatin 4 is a nanomolar inhibitor of the food vacuole falcipains. *Chemistry and Biology.* 19, 1546-55.
157. Li H, Ponder EL, Verdoes M, Asbjornsdottir KH, Deu E, Edgington LE, Lee JT, Kirk CJ, Demo SD, Williamson KC, **Bogyo M**. (2012) Validation of the proteasome as a therapeutic target in Plasmodium using an epoxyketone inhibitor with parasite-specific toxicity. *Chemistry and Biology.* 19, 1535-45.

158. Edgington LE, Verdoes M, Ortega A, Withana NP, Lee J, Syed S, Bachmann MH, Blum G, **Bogyo M**. (2013) Functional Imaging of Legumain in Cancer Using a New Quenched Activity-Based Probe. *J Am Chem Soc*. 135, 174-82
159. Mullins SR, Sameni M, Blum G, **Bogyo M**, Sloane B, Moin K. (2013) Three-dimensional cultures modeling premalignant progression of human breast epithelial cells: role of cysteine cathepsins. *Biol Chem*. 393, 1405-16.
160. Edgington, LE and **Bogyo, M**. (2013) In Vivo Imaging and Biochemical Characterization of Protease Function Using Fluorescent Activity-Based Probes *Current Protocols in Chemical Biology* 5, 1-20.
161. Lu H, Wang Z, Shabab M, Oeljeklaus J, Verhelst SH, Kaschani F, Kaiser M, **Bogyo M**, van der Hoorn RA. (2013) A substrate-inspired probe monitors translocation, activation, and subcellular targeting of bacterial type III effector protease AvrPphB. *Chem Biol*. 20, 168-76.
162. Lee J, **Bogyo M**. (2013) Target deconvolution techniques in modern phenotypic profiling. *Curr Opin Chem Biol*. 17, 118-26.
163. Xiao J, Broz P, Puri AW, Deu E, Morell M, Monack DM, **Bogyo M**. (2013) A coupled protein and probe engineering approach for selective inhibition and activity-based probe labeling of the caspases. *J Am Chem Soc* 135, 9130-8.
164. Morell M, Nguyen Duc T, Willis AL, Syed S, Lee J, Deu E, Deng Y, Xiao J, Turk BE, Jessen JR, Weiss SJ, **Bogyo M**. (2013) Coupling protein engineering with probe design to inhibit and image matrix metalloproteinases with controlled specificity. *J Am Chem Soc*. 135, 9139-48.
165. Godinat A, Park HM, Miller SC, Cheng K, Hanahan D, Sanman LE, **Bogyo M**, Yu A, Nikitin GF, Stahl A, Dubikovskaya EA. (2013) A biocompatible *in vivo* ligation reaction and its application for noninvasive bioluminescent imaging of protease activity in living mice. *ACS Chem Biol*. 8, 987-99.
166. Tanaka TQ, Deu E, Molina-Cruz A, Ashburne MF, Ali O, Suri A, Kortagere S, **Bogyo M**, Williamson KC. (2013) Plasmodium dipeptidyl aminopeptidases as malaria transmission blocking drug targets. *Antimicrob Agents Chemother*. 57, 4645-52.
167. Gloeckl S, Ong VA, Patel P, Tyndall JD, Timms P, Beagley KW, Allan JA, Armitage CW, Turnbull L, Whitchurch CB, Merdanovic M, Ehrmann M, Powers JC, Oleksyszyn J, Verdoes M, **Bogyo M**, Huston WM. (2013) Identification of a serine protease inhibitor which causes inclusion vacuole reduction and is lethal to *Chlamydia trachomatis*. *Mol Microbiol*. 89, 676-89.
168. Child, MA, Hall, CI, Beck, JR, Ofori, LO, Albrow, VE, Garland, M, Bowyer, PW, Bradley, PJ, Powers, JC, Boothroyd, JC, Weerapana, E, **Bogyo, M**. (2013) Small-molecule inhibition of a depalmitoylase enhances *Toxoplasma* host cell invasion. *Nat. Chem. Bio*. 9, 651-6.
169. Verdoes M, Oresic Bender K, Segal E, van der Linden WA, Syed S, Withana NP, Sanman LE, **Bogyo M**. (2013) An improved quenched fluorescent probe for imaging of cysteine cathepsin activity. *J Am Chem Soc*. 135, 14726-30.
170. Ruffell B, Affara NI, Cottone L, Junankar S, Johansson M, Denardo DG, Korets L, Reinheckel T, Sloane BF, **Bogyo M**, Coussens LM. (2013) Cathepsin C is a tissue-specific regulator of squamous carcinogenesis. *Genes Dev*. 27, 2086-98.
171. Deu E, Chen IT, Lauterwasser EM, Valderramos J, Li H, Edgington LE, Renslo AR, **Bogyo M**. (2013) Ferrous iron-dependent drug delivery enables controlled and selective release of therapeutic agents in vivo. *Proc Natl Acad Sci U S A*. 110, 18244-9.
172. Rothberg JM, Bailey KM, Wojtkowiak JW, Ben-Nun Y, **Bogyo M**, Weber E, Moin K, Blum G, Mattingly RR, Gillies RJ, Sloane BF (2013) Acid-mediated tumor proteolysis: contribution of cysteine cathepsins. *Neoplasia*. 15, 1125-37.
173. Puri AW, **Bogyo M**. (2013) Applications of Small Molecule Probes in Dissecting Mechanisms of Bacterial Virulence and Host Responses. *Biochemistry*. 52, 5985-96.

174. Rothberg JM, Bailey KM, Wojtkowiak JW, Ben-Nun Y, **Bogyo M**, Weber E, Moin K, Blum G, Mattingly RR, Gillies RJ, Sloane BF. (2013) Acid-mediated tumor proteolysis: contribution of cysteine cathepsins. *Neoplasia*. 15, 1125-37.
175. Razorenova OV, Castellini L, Colavitti R, Edgington LE, Nicolau M, Huang X, Bedogni B, Mills EM, **Bogyo M**, Giaccia AJ. (2014). The apoptosis repressor with a CARD domain (ARC) gene is a direct hypoxia-inducible factor 1 target gene and promotes survival and proliferation of VHL-deficient renal cancer cells. *Mol Cell Biol*. 34, 739-51.
176. Walker E, Gopalakrishnan R, **Bogyo M**, Basilion JP. (2014) Microscopic detection of quenched activity-based optical imaging probes using an antibody detection system: localizing protease activity. *Mol Imaging Biol*. 16, 608-18.
177. Cattaruzza F, Amadesi S, Carlsson JF, Murphy JE, Lyo V, Kirkwood K, Cottrell GS, Bogyo M, Knecht W, Bunnett NW. (2014). Serine proteases and protease-activated receptor 2 mediate the proinflammatory and algescic actions of diverse stimulants. *Br J Pharmacol*. 171, 3814-26.
178. Haedke UR, Frommel SC, Hansen F, Hahne H, Kuster B, Bogyo M, Verhelst SH. (2014). Phosphoramidates as novel activity-based probes for serine proteases. *Chembiochem*. 15, 1106-10.
179. Sanman LE, Bogyo M. Activity-based profiling of proteases. (2014) *Annu Rev Biochem*. 83, 249-73.
180. Li H, van der Linden WA, Verdoes M, Florea BI, McAllister FE, Govindaswamy K, Elias JE, Bhanot P, Overkleeft HS, Bogyo M. (2014). Assessing subunit dependency of the Plasmodium proteasome using small molecule inhibitors and active site probes. *ACS Chem Biol*. 9, 1869-76.
181. Gangoda L, Doerflinger M, Srivastava R, Narayan N, Edgington LE, Orian J, Hawkins C, O'Reilly LA, Gu H, Bogyo M, Ekert P, Strasser A, Puthalakath H. (2014). Loss of Prkar1a leads to Bcl-2 family protein induction and cachexia in mice. *Cell Death Differ*. 21, 1815-24.
182. Godinat A, Budin G, Morales AR, Park HM, Sanman LE, Bogyo M, Yu A, Stahl A, Dubikovskaya EA. (2014). A biocompatible "split luciferin" reaction and its application for non-invasive bioluminescent imaging of protease activity in living animals. *Curr Protoc Chem Biol*. 6, 169-89.
183. Li H, Tsu C, Blackburn C, Li G, Hales P, Dick L, **Bogyo M**. (2014) Identification of Potent and Selective Non-covalent Inhibitors of the Plasmodium falciparum Proteasome. *J Am Chem Soc*. 136, 13562-5.
184. Segal E, Prestwood TR, van der Linden WA, Carmi Y, Bhattacharya N, Withana N, Verdoes M, Habtezion A, Engleman EG, **Bogyo M**. (2015) Detection of intestinal cancer by local, topical application of a quenched fluorescence probe for cysteine cathepsins. *Chem Biol*. 22, 148-58.
185. Hachmann J, Edgington-Mitchell LE, Poreba M, Sanman LE, Drag M, **Bogyo M**, Salvesen GS. (2015) Probes to monitor activity of the paracaspase MALT1. *Chem Biol*. 22, 139-47.
186. Oresic Bender K, Ofori L, van der Linden WA, Mock ED, Datta GK, Chowdhury S, Li H, Segal E, Sanchez Lopez M, Ellman JA, Figdor CG, **Bogyo M**, Verdoes M. (2015) Design of a Highly Selective Quenched Activity-Based Probe and Its Application in Dual Color Imaging Studies of Cathepsin S Activity Localization. *J Am Chem Soc*. 137, 4771-7.
187. Gangoda L, Keerthikumar S, Fonseka P, Edgington LE, Ang CS, Ozcitti C, **Bogyo M**, Parker BS, Mathivanan S. (2015). Inhibition of cathepsin proteases attenuates migration and sensitizes aggressive N-Myc amplified human neuroblastoma cells to doxorubicin. *Oncotarget*. 6, 11175-90.
188. Ofori LO, Withana NP, Prestwood TR, Verdoes M, Brady JJ, Winslow MM, Sorger J, **Bogyo M**. (2015) Design of Protease Activated Optical Contrast Agents That Exploit a Latent Lysosomotropic Effect for Use in Fluorescence-Guided Surgery. *ACS Chem Biol*. 10, 1977-88.

189. Lu H, Chandrasekar B, Oeljeklaus J, Misas-Villamil JC, Wang Z, Shindo T, **Bogyo M**, Kaiser M, van der Hoorn RA. (2015) Subfamily-specific Fluorescent Probes for Cys proteases Display Dynamic Protease Activities During Seed Germination. *Plant Physiol.* 168, 1462-75.
190. Orłowski GM, Colbert JD, Sharma S, **Bogyo M**, Robertson SA, Rock KL. (2015) Multiple Cathepsins Promote Pro-IL-1 β Synthesis and NLRP3-Mediated IL-1 β Activation. *J Immunol.* 195, 1685-97.
191. van der Linden WA, Segal E, Child MA, Byzia A, Drag M, **Bogyo M**. (2015) Design and Synthesis of Activity-Based Probes and Inhibitors for Bleomycin Hydrolase. *Chem Biol.* 22, 995-1001.
192. Edgington-Mitchell LE, Rautela J, Duivenvoorden HM, Jayatilleke KM, van der Linden WA, Verdoes M, **Bogyo M**, Parker BS. (2015) Cysteine cathepsin activity suppresses osteoclastogenesis of myeloid-derived suppressor cells in breast cancer. *Oncotarget.* 6, 27008-22.
193. Hou L, Cooley J, Swanson R, Ong PC, Pike RN, **Bogyo M**, Olson ST, Remold-O'Donnell E. (2015) The protease cathepsin L regulates Th17 cell differentiation. *J Autoimmun.* 65, 56-63.
194. Staniec D, Ksiazek M, Thøgersen IB, Enghild JJ, Sroka A, Bryzek D, **Bogyo M**, Abrahamson M, Potempa J. (2015) Calcium Regulates the Activity and Structural Stability of Tpr, a Bacterial Calpain-like Peptidase. *J Biol Chem.* 290, 27248-60.
195. Bender KO, Garland M, Ferreyra JA, Hryckowian AJ, Child MA, Puri AW, Solow-Cordero DE, Higginbottom SK, Segal E, Banaei N, Shen A, Sonnenburg JL, **Bogyo M**. (2015) A small-molecule antivirulence agent for treating *Clostridium difficile* infection. *Sci Transl Med.* 7, 306ra148.
196. Rosenthal EL, Warram JM, de Boer E, Basilion JP, Biel MA, **Bogyo M**, Bouvet M, Brigman BE, Colson YL, DeMeester SR, Gurtner GC, Ishizawa T, Jacobs PM, Keereweer S, Liao JC, Nguyen QT, Olson JM, Paulsen KD, Rieves D, Sumer BD, Tweedle MF, Vahrmeijer AL, Weichert JP, Wilson BC, Zenn MR, Zinn KR, van Dam GM. (2015) Successful Translation of Fluorescence Navigation During Oncologic Surgery: A Consensus Report. *J Nucl Med.* 57, 144-50.
197. Foe IT, Child MA, Majmudar JD, Krishnamurthy S, van der Linden WA, Ward GE, Martin BR, **Bogyo M**. (2015) Global Analysis of Palmitoylated Proteins in *Toxoplasma gondii*. *Cell Host Microbe.* 18, 501-11.
198. Lauterwasser EM, Fontaine SD, Li H, Gut J, Katneni K, Charman SA, Rosenthal PJ, **Bogyo M**, Renslo AR. (2015) Trioxolane-Mediated Delivery of Mefloquine Limits Brain Exposure in a Mouse Model of Malaria. *ACS Med Chem Lett.* 6, 1145-9.
199. Withana NP, Garland M, Verdoes M, Ofori LO, Segal E, **Bogyo M**. (2016) Labeling of active proteases in fresh-frozen tissues by topical application of quenched activity-based probes. *Nature Protoc.* 1, 184-91.
200. Withana NP, Ma X, McGuire HM, Verdoes M, van der Linden WA, Ofori LO, Zhang R, Li H, Sanman LE, Wei K, Yao S, Wu P, Li F, Huang H, Xu Z, Wolters PJ, Rosen GD, Collard HR, Zhu Z, Cheng Z, **Bogyo M**. (2016) Non-invasive Imaging of Idiopathic Pulmonary Fibrosis Using Cathepsin Protease Probes. *Sci Rep.* 6, 19755.
201. Li, H., O'Donoghue, A.J., van der Linden, W.A., Xie, S.C., Yoo, E., Too, E., Foe, I.T., Tilley, L., Craik, C.S., da Fonseca, P.C.A, **Bogyo, M**. (2016) Structure and function-based design of Plasmodium-selective proteasome inhibitors. *Nature.* 530, 233-6.
202. Garland M, Yim JJ, **Bogyo M**. (2016) A Bright Future for Precision Medicine: Advances in Fluorescent Chemical Probe Design and Their Clinical Application. *Cell Chem Biol.* 23, 122-36.
203. Sanman LE, Qian Y, Eisele NA, Ng TM, van der Linden WA, Monack DM, Weerapana E, **Bogyo M**. (2016) Disruption of glycolytic flux is a signal for inflammasome signaling and pyroptotic cell death. *Elife.* 5, e13663.
204. Edgington-Mitchell LE, **Bogyo M**. (2016) Detection of Active Caspases During Apoptosis Using Fluorescent Activity-Based Probes. *Methods Mol Biol.* 1419, 27-39.

205. Sensarn S, Zavaleta CL, Segal E, Rogalla S, Lee W, Gambhir SS, **Bogyo M**, Contag CH. (2016) A Clinical Wide-Field Fluorescence Endoscopic Device for Molecular Imaging Demonstrating Cathepsin Protease Activity in Colon Cancer. *Mol Imaging Biol.* 18, 820-829.
206. Withana NP, Saito T, Ma X, Garland M, Liu C, Kosuge H, Amsallem M, Verdoes M, Ofori LO, Fischbein M, Arakawa M, Cheng Z, McConnell MV, **Bogyo M**. (2016) Dual Modality Activity Based Probes as Molecular Imaging Agents for Vascular Inflammation. *J Nucl Med* 57, 1583-1590.
207. Li H, **Bogyo M**, da Fonseca PC. (2016) The cryo-EM structure of the Plasmodium falciparum 20S proteasome and its use in the fight against malaria. *FEBS J.* Jun 11. doi: 10.1111/febs.13780.
208. van der Linden WA, Schulze CJ, Herbert AS, Krause TB, Wirchnianski AA, Dye JM, Chandran K, **Bogyo M**. (2016) Cysteine Cathepsin Inhibitors as Anti-Ebola Agents. *ACS Infect Dis.* 2, 173-179.
209. Sanman LE, van der Linden WA, Verdoes M, **Bogyo M**. (2016) Bifunctional Probes of Cathepsin Protease Activity and pH Reveal Alterations in Endolysosomal pH during Bacterial Infection. *Cell Chem Biol.* 23, 793-804.
210. Edgington-Mitchell LE, Wartmann T, Fleming AK, Gocheva V, van der Linden WA, Withana NP, Verdoes M, Aurelio L, Edgington-Mitchell D, Lieu T, Parker BS, Graham B, Reinheckel T, Furness JB, Joyce JA, Storz P, Halangk W, **Bogyo M**, Bunnett NW. (2016) Legumain is Activated in Macrophages during Pancreatitis. *Am J Physiol Gastrointest Liver Physiol.* 311, G548-60.
211. Abd-Elrahman I, Kosuge H, Wisnes Sadan T, Ben-Nun Y, Meir K, Rubinstein C, **Bogyo M**, McConnell MV, Blum G. (2016) Cathepsin Activity-Based Probes and Inhibitor for Preclinical Atherosclerosis Imaging and Macrophage Depletion. *PLoS One.* 11, e0160522.
212. Grüner BM, Schulze CJ, Yang D, Ogasawara D, Dix MM, Rogers ZN, Chuang CH, McFarland CD, Chiou SH, Brown JM, Cravatt BF, **Bogyo M***, Winslow MM*. (2016) An in vivo multiplexed small-molecule screening platform. *Nat Methods.* 13, 883-9.
213. Lentz CS, Ordonez AA, Kasperkiewicz P, La Greca F, O'Donoghue AJ, Schulze CJ, Powers JC, Craik CS, Drag M, Jain SK, **Bogyo M**. (2016) Design of Selective Substrates and Activity-Based Probes for Hydrolase Important for Pathogenesis 1 (HIP1) from Mycobacterium tuberculosis. *ACS Infect Dis.* 2, 807-81.
214. Edgington-Mitchell LE, **Bogyo M**, Verdoes M. (2016). Live Cell Imaging and Profiling of Cysteine Cathepsin Activity Using a Quenched Activity-Based Probe. *Methods Mol Biol.* 1491, 145-159.
215. Sensarn S, Zavaleta CL, Segal E, Rogalla S, Lee W, Gambhir SS, **Bogyo M**, Contag CH. (2016) A Clinical Wide-Field Fluorescence Endoscopic Device for Molecular Imaging Demonstrating Cathepsin Protease Activity in Colon Cancer. *Mol Imaging Biol.* 18, 820-829.
216. Dubey R, Staker BL, Foe IT, **Bogyo M**, Myler PJ, Ngô HM, Gubbels MJ. (2017) Membrane skeletal association and post-translational allosteric regulation of Toxoplasma gondii GAPDH1 *Mol Microbiol.* 103, 618-634.
217. Child MA, Garland M, Foe I, Madzelan P, Treeck M, van der Linden WA, Oresic Bender K, Weerapana E, Wilson MA, Boothroyd JC, Reese ML, **Bogyo M**. (2017) Toxoplasma DJ-1 Regulates Organelle Secretion by a Direct Interaction with Calcium-Dependent Protein Kinase 1. *MBio.* 8(1). pii: e02189-16.
218. Walker E, Mann M, Honda K, Vidimos A, Schluchter MD, Straight B, **Bogyo M**, Popkin D, Basilion JP. (2017) Rapid visualization of nonmelanoma skin cancer. *J Am Acad Dermatol.* 76, 209-216.
219. Bender KO, Garland M, **Bogyo M**. (2017) Response to Comment on "A small-molecule antivirulence agent for treating Clostridium difficile infection". *Sci Transl Med.* 8, 370tr2.
220. Orłowski GM, Sharma S, Colbert JD, **Bogyo M**, Robertson SA, Kataoka H, Chan FK, Rock KL. (2017) Frontline Science: Multiple cathepsins promote inflammasome-independent, particle-induced cell death during NLRP3-dependent IL-1 β activation. *J Leukoc Biol.* 102, 7-17.

221. Hewings DS, Flygare JA, Wertz IE, **Bogyo M.** (2017) Activity-based probes for the multicatalytic proteasome. *FEBS J.* 284, 1540-1554.
222. Tipirneni KE, Rosenthal EL, Moore LS, Haskins AD, Udayakumar N, Jani AH, Carroll WR, Morlandt AB, **Bogyo M,** Rao J, Warram JM. (2017) Fluorescence Imaging for Cancer Screening and Surveillance. *Mol Imaging Biol.* 19, 645-655.
223. Hewings DS, Flygare JA, **Bogyo M,** Wertz IE. (2017) Activity-based probes for the ubiquitin conjugation-deconjugation machinery: new chemistries, new tools, and new insights. *FEBS J.* 284(10):1555-1576.
224. Garland M, Loscher S, **Bogyo M.** (2017) Chemical Strategies To Target Bacterial Virulence. *Chem Rev.* 117, 4422-4461.
225. Ward ME, Chen R, Huang HY, Ludwig C, Telpoukhovskaia M, Taubes A, Boudin H, Minami SS, Reichert M, Albrecht P, Gelfand JM, Cruz-Herranz A, Cordano C, Alavi MV, Leslie S, Seeley WW, Miller BL, Bigio E, Mesulam MM, **Bogyo M,** Mackenzie IR, Staropoli JF, Cotman SL, Huang EJ, Gan L, Green AJ. (2017) Individuals with progranulin haploinsufficiency exhibit features of neuronal ceroid lipofuscinosis. *Sci Transl Med.* 12, 9(385).
226. Di Cristina M, Dou Z, Lunghi M, Kannan G, Huynh MH, McGovern OL, Schultz TL, Schultz AJ, Miller AJ, Hayes BM, van der Linden W, Emiliani C, **Bogyo M,** Besteiro S, Coppens I, Carruthers VB. (2017) Toxoplasma depends on lysosomal consumption of autophagosomes for persistent infection. *Nat Microbiol.* 2,17096.
227. Ng CL, Fidock DA, **Bogyo M.** Protein Degradation Systems as Antimalarial Therapeutic Targets. (2017) *Trends Parasitol.* 33, 731-743.
228. Lee CW, Stankowski JN, Chew J, Cook CN, Lam YW, Almeida S, Carlomagno Y, Lau KF, Prudencio M, Gao FB, **Bogyo M,** Dickson DW, Petrucelli L. (2017) The lysosomal protein cathepsin L is a progranulin protease. *Mol Neurodegener.* 12, 55.
229. Hopp CS, Bennett BL, Mishra S, Lehmann C, Hanson KK, Lin JW, Rousseau K, Carvalho FA, van der Linden WA, Santos NC, **Bogyo M,** Khan SM, Heussler V, Sinnis P. (2017) Deletion of the rodent malaria ortholog for falcipain-1 highlights differences between hepatic and blood stage merozoites. *PLoS Pathog.* 13, e1006586.
230. Duivenvoorden HM, Rautela J, Edgington-Mitchell LE, Spurling A, Greening DW, Nowell CJ, Molloy TJ, Robbins E, Brockwell NK, Lee CS, Chen M, Holliday A, Selinger CI, Hu M, Britt KL, Stroud DA, **Bogyo M,** Möller A, Polyak K, Sloane BF, O'Toole SA, Parker BS. (2017) Myoepithelial cell-specific expression of stefin A as a suppressor of early breast cancer invasion. *J Pathol.* 243, 496-509.
231. Aillaud C, Bosc C, Peris L, Bosson A, Heemeryck P, Van Dijk J, Le Fric J, Boulan B, Vossier F, Sanman LE, Syed S, Amara N, Couté Y, Lafanechère L, Denarier E, Delphin C, Pelletier L, Humbert S, **Bogyo M,** Andrieux A, Rogowski K, Moutin MJ. (2017) Vasohibins/SVBP are tubulin carboxypeptidases (TCPs) that regulate neuron differentiation. *Science.* 358, 1448-1453.
232. Yim JJ, Tholen M, Klaassen A, Sorger J, **Bogyo M.** (2018) Optimization of a Protease Activated Probe for Optical Surgical Navigation. *Mol Pharm.* 15, 750-758.
233. Tomlin FM, Gerling-Driessen UIM, Liu YC, Flynn RA, Vangala JR, Lentz CS, Clauder-Muenster S, Jakob P, Mueller WF, Ordoñez-Rueda D, Paulsen M, Matsui N, Foley D, Rafalko A, Suzuki T, **Bogyo M,** Steinmetz LM, Radhakrishnan SK, Bertozzi CR. (2018) Inhibition of NGLY1 Inactivates the Transcription Factor Nrf1 and Potentiates Proteasome Inhibitor Cytotoxicity. *ACS Cent Sci.* 3, 1143-1155.
234. Garland M, Schulze CJ, Foe IT, van der Linden WA, Child MA, **Bogyo M.** (2018) Development of an activity-based probe for acyl-protein thioesterases. *PLoS One.* 13(1), e0190255.
235. **Bogyo M,** Yim JJ, Rosenthal EL. (2018) New Blood Test SEEKs To Detect and Localize Cancer before It's Too Late. *Biochemistry.* 57, 1561-1562.

236. Flanagan-Steet H, Christian C, Lu PN, Aarnio-Peterson M, Sanman L, Archer-Hartmann S, Azadi P, **Bogyo M**, Steet RA. (2018) TGF- β Regulates Cathepsin Activation during Normal and Pathogenic Development. *Cell Rep.* 22, 2964-2977.
237. Hewings DS, Heideker J, Ma TP, AhYoung AP, El Oualid F, Amore A, Costakes GT, Kirchhofer D, Brasher B, Pillow T, Popovych N, Maurer T, Schwerdtfeger C, Forrest WF, Yu K, Flygare J, **Bogyo M**, Wertz IE. (2018) Reactive-site-centric chemoproteomics identifies a distinct class of deubiquitinase enzymes. *Nat Commun.* 9, 1162.
238. Lentz CS, Sheldon JR, Crawford LA, Cooper R, Garland M, Amieva MR, Weerapana E, Skaar EP, **Bogyo M**. Identification of a *S. aureus* virulence factor by activity-based protein profiling (ABPP). (2018) *Nat Chem Biol.* 14, 609.
239. Stathopoulou C, Gangaplara A, Mallett G, Flomerfelt FA, Liniany LP, Knight D, Samsel LA, Berlinguer-Palmini R, Yim JJ, Felizardo TC, Eckhaus MA, Edgington-Mitchell L, Martinez-Fabregas J, Zhu J, Fowler DH, van Kasteren SI, Laurence A, **Bogyo M**, Watts C, Shevach EM, Amarnath S. PD-1 Inhibitory Receptor Downregulates Asparaginyl Endopeptidase and Maintains Foxp3 Transcription Factor Stability in Induced Regulatory T Cells. *Immunity.* (2018) 49, 247-263.
240. Yoo E, Stokes BH, de Jong H, Vanaerschot M, Kumar T, Lawrence N, Njoroge M, Garcia A, Van der Westhuyzen R, Momper JD, Ng CL, Fidock DA, **Bogyo M**. Defining the Determinants of Specificity of Plasmodium Proteasome Inhibitors. *J Am Chem Soc.* (2018) 140, 11424-11437.
241. Amara N, Tholen M, **Bogyo M**. Chemical Tools for Selective Activity Profiling of Endogenously Expressed MMP-14 in Multicellular Models. *ACS Chem Biol.* (2018) 13, 2645-2654.
242. Foe IT, Onguka O, Amberg-Johnson K, Garner RM, Amara N, Beatty W, Yeh E, **Bogyo M**. The *Toxoplasma gondii* Active Serine Hydrolase 4 Regulates Parasite Division and Intravacuolar Parasite Architecture. *mSphere.* (2018) 3(5). pii: e00393-18.
243. Seth-Pasricha M, Senn S, Sanman LE, **Bogyo M**, Nanda V, Bidle KA, Bidle KD. Catalytic linkage between caspase activity and proteostasis in Archaea. *Environ Microbiol.* 2018 Oct 28. doi: 10.1111/1462-2920.14456.
244. Shrivastav M, Gounaris E, Khan MW, Ko J, Ryu SH, **Bogyo M**, Larson A, Barret TA, Bentrem DJ. Validation of near infrared fluorescence (NIRF) probes in vivo with dual laser NIRF endoscope. *PLoS One.* 2018 Nov 2;13(11):e0206568.
245. Amara N, Foe, IT, Onguka, O, Garland M, **Bogyo M**. Synthetic fluorogenic peptides reveal dynamic substrate specificity of depalmitoylases. *Cell Chem Bio* 2018 Oct 22. pii: S2451-9456(18)30338-6.
246. Garland M, Babin BM, Miyashita SI, Loscher S, Shen Y, Dong M, **Bogyo M**. Covalent modifiers of Botulinum neurotoxin counteract toxin persistence. *ACS Chem Biol.* 2019 14(1):76-87.
247. Chen L, Keller LJ, Cordasco E, **Bogyo M**, Lentz CS. Fluorescent Triazole Urea Activity-Based Probes for the Single-Cell Phenotypic Characterization of *Staphylococcus aureus*. *Angew Chem Int Ed Engl.* 2019 58:5643-5647.
248. Liu Y, Walker E, Iyer SR, Biro M, Kim I, Zhou B, Straight B, **Bogyo M**, Basilion JP, Popkin DL, Wilson DL. Molecular imaging and validation of margins in surgically excised nonmelanoma skin cancer specimens. *J Med Imaging (Bellingham).* 2019 6:016001.
249. Stokes BH, Yoo E, Murithi JM, Luth MR, Afanasyev P, da Fonseca PCA, Winzeler EA, Ng CL, Bogyo M, Fidock DA. Covalent *Plasmodium falciparum*-selective proteasome inhibitors exhibit a low propensity for generating resistance in vitro and synergize with multiple antimalarial agents. *PLoS Pathog.* 2019 Jun 6;15(6):e1007722.
250. de Vries LE, Sanchez MI, Groborz K, Kuppens L, Poreba M, Lehmann C, Nevins N, Withers-Martinez C, Hirst DJ, Yuan F, Arastu-Kapur S, Horn M, Mares M, **Bogyo M**, Drag M, Deu E. Characterization of *P. falciparum* dipeptidyl aminopeptidase 3 specificity identifies differences in amino acid preferences between peptide-based substrates and covalent inhibitors. *FEBS J.* 2019 286:3998-4023.

251. Veillard F, Sztukowska M, Nowakowska Z, Mizgalska D, Thøgersen IB, Enghild JJ, **Bogyo M**, Potempa B, Nguyen KA, Potempa J. Proteolytic processing and activation of gingipain zymogens secreted by T9SS of *Porphyromonas gingivalis*. *Biochimie*. 2019 166:161-172.
252. Szumska J, Batool Z, Al-Hashimi A, Venugopalan V, Skripnik V, Schaschke N, **Bogyo M**, Brix K. Treatment of rat thyrocytes in vitro with cathepsin B and L inhibitors results in disruption of primary cilia leading to redistribution of the trace amine associated receptor 1 to the endoplasmic reticulum. *Biochimie*. 2019 166:270-285.
253. Babin BM, Kasperkiewicz P, Janiszewski T, Yoo E, Drag M, **Bogyo M**. Leveraging peptide substrate libraries to design inhibitors of bacterial Lon protease. *ACS Chem Biol*. 2019 14, 2453-2462.
254. Chen S, Yim JJ, **Bogyo M**. Synthetic and biological approaches to map substrate specificities of proteases. *Biol Chem*. 2019 401(1):165-182.
255. Keller LJ, Babin BM, Lakemeyer M, **Bogyo M**. Activity-based protein profiling in bacteria: Applications for identification of therapeutic targets and characterization of microbial communities. *Curr Opin Chem Biol*. 2020 54:45-53.
256. Sanchez MI, de Vries LE, Lehmann C, Lee JT, Ang KK, Wilson C, Chen S, Arkin MR, **Bogyo M**, Deu E. Identification of Plasmodium dipeptidyl aminopeptidase allosteric inhibitors by high throughput screening. *PLoS One*. 2019 Dec 18;14(12):e0226270.
257. Yoo E, Schulze CJ, Stokes BH, Onguka O, Yeo T, Mok S, Gnädig NF, Zhou Y, Kurita K, Foe IT, Terrell SM, Boucher MJ, Cieplak P, Kumpornsin K, Lee MCS, Lington RG, Long JZ, Uhlemann AC, Weerapana E, Fidock DA, **Bogyo M**. The Antimalarial Natural Product Salinipostin A Identifies Essential α/β Serine Hydrolases Involved in Lipid Metabolism in *P. falciparum* Parasites. *Cell Chem Biol*. 2020 Feb 20;27(2):143-157
258. Walker E, Liu Y, Kim I, Biro M, Iyer SR, Ezaldein H, Scott J, Merati M, Mistur R, Zhou B, Straight B, Yim JJ, **Bogyo M**, Mann M, Wilson DL, Basilion JP, Popkin DL. A Protease-Activated Fluorescent Probe Allows Rapid Visualization of Keratinocyte Carcinoma During Excision. *Cancer Res*. 2020 pii: canres.3067.2019.
259. Keller LJ, Lentz CS, Chen YE, Metivier RJ, Weerapana E, Fischbach MA, **Bogyo M**. Characterization of Serine Hydrolases Across Clinical Isolates of Commensal Skin Bacteria *Staphylococcus epidermidis* Using Activity-Based Protein Profiling. *ACS Infect Dis*. 2020 May 8;6(5):930-938.
260. Widen JC, Tholen M, Yim JJ, **Bogyo M**. Methods for analysis of near-infrared (NIR) quenched-fluorescent contrast agents in mouse models of cancer. *Methods Enzymol*. 2020;639:141-166.
261. Garland M, Hryckowian AJ, Tholen M, Bender KO, Van Treuren WW, Loscher S, Sonnenburg JL, **Bogyo M**. The Clinical Drug Ebselen Attenuates Inflammation and Promotes Microbiome Recovery in Mice after Antibiotic Treatment for CDI. *Cell Rep Med*. 2020 Apr 21;1(1):100005.
262. Tholen M, Yim JJ, Groborz K, Yoo E, Martin BA, van den Berg NS, Drag M, **Bogyo M**. Design of optical imaging probes by screening of diverse substrate libraries directly in disease tissue extracts. *Angew Chem Int Ed Engl*. 2020 59(43):19143-19152.
263. Faucher F, Bennett JM, **Bogyo M**, Lovell S. Strategies for Tuning the Selectivity of Chemical Probes that Target Serine Hydrolases. *Cell Chem Biol*. 2020 27(8):937-952.
264. Telpoukhovskaia MA, Liu K, Sayed FA, Etchegaray JI, Xie M, Zhan L, Li Y, Zhou Y, Le D, Bahr BA, **Bogyo M**, Ding S, Gan L. Discovery of small molecules that normalize the transcriptome and enhance cysteine cathepsin activity in progranulin-deficient microglia. *Sci Rep*. 2020 10(1):13688.

265. Fellner M, Lentz CS, Jamieson SA, Brewster JL, Chen L, **Bogyo M**, Mace PD. Structural Basis for the Inhibitor and Substrate Specificity of the Unique Fph Serine Hydrolases of *Staphylococcus aureus*. *ACS Infect Dis*. 2020 6(10):2771-2782.
266. Bilverstone TW, Garland M, Cave RJ, Kelly ML, Tholen M, Bouley DM, Kaye P, Minton NP, **Bogyo M**, Kuehne SA, Melnyk RA. The glucosyltransferase activity of *C. difficile* Toxin B is required for disease pathogenesis. *PLoS Pathog* 2020 Sep 22;16(9):e1008852.
267. Widen JC, Tholen M, Yim JJ, Antaris A, Casey KM, Rogalla S, Klaassen A, Sorger J, **Bogyo M**. AND-gate contrast agents for enhanced fluorescence-guided surgery. *Nat Biomed Eng*. 2020 5(3):264-277.
268. Suurs FV, Qiu SQ, Yim JJ, Schröder CP, Timmer-Bosscha H, Bensen ES, Santini JT Jr, de Vries EGE, **Bogyo M**, van Dam GM. Fluorescent image-guided surgery in breast cancer by intravenous application of a quenched fluorescence activity-based probe for cysteine cathepsins in a syngeneic mouse model. *EJNMMI Res*. 2020 Sep 29;10(1):111.
269. Simwela NV, Stokes BH, Aghabi D, **Bogyo M**, Fidock DA, Waters AP. Plasmodium berghei K13 Mutations Mediate In Vivo Artemisinin Resistance That Is Reversed by Proteasome Inhibition. *mBio*. 2020 Nov 10;11(6):e02312-20.
270. Yim JJ, Singh SP, Xia A, Kashfi-Sadabad R, Tholen M, Huland DM, Zarabanda D, Cao Z, Solis-Pazmino P, **Bogyo M**, Valdez TA. Short-Wave Infrared Fluorescence Chemical Sensor for Detection of Otitis Media. *ACS Sens*. 2020 Nov 25;5(11):3411-3419.
271. Chen S, Lovell S, Lee S, Fellner M, Mace PD, **Bogyo M**. Identification of highly selective covalent inhibitors by phage display. *Nat Biotechnol*. 2020 Apr;39(4):490-498.
272. Brodzicki A, Jaworek-Korjakowska J, Kleczek P, Garland M, **Bogyo M**. Pre-Trained Deep Convolutional Neural Network for *Clostridioides Difficile* Bacteria Cytotoxicity Classification Based on Fluorescence Images. *Sensors (Basel)*. 2020 20(23):6713.
273. Al-Hashimi A, Venugopalan V, Rehders M, Sereesongsang N, Hein Z, Springer S, Weber E, Führer D, **Bogyo MS**, Scott CJ, Burden RE, Brix K. Procathepsin V Is Secreted in a TSH Regulated Manner from Human Thyroid Epithelial Cells and Is Accessible to an Activity-Based Probe. *Int J Mol Sci*. 2020 Nov 30;21(23):9140.
274. Yim JJ, Harmsen S, Flisikowski K, Flisikowska T, Namkoong H, Garland M, van den Berg NS, Vilches-Moure JG, Schnieke A, Saur D, Glasl S, Gorpas D, Habtezion A, Ntziachristos V, Contag CH, Gambhir SS, **Bogyo M**, Rogalla S. A protease-activated, near-infrared fluorescent probe for early endoscopic detection of premalignant gastrointestinal lesions. *Proc Natl Acad Sci U S A*. 2021 Jan 5;118(1):e2008072118.
275. Venugopalan V, Al-Hashimi A, Rehders M, Golchert J, Reinecke V, Homuth G, Völker U, Manirajah M, Touzani A, Weber J, **Bogyo MS**, Verrey F, Wirth EK, Schweizer U, Heuer H, Kirstein J, Brix K. The Thyroid Hormone Transporter Mct8 Restricts Cathepsin-Mediated Thyroglobulin Processing in Male Mice through Thyroid Auto-Regulatory Mechanisms That Encompass Autophagy. *Int J Mol Sci*. 2021 Jan 5;22(1):462.
276. Steuten K, Kim H, Widen JC, Babin BM, Onguka O, Lovell S, Bolgi O, Cerikan B, Neufeldt CJ, Cortese M, Muir RK, Bennett JM, Geiss-Friedlander R, Peters C, Bartenschlager R, **Bogyo M**. Challenges for Targeting SARS-CoV-2 Proteases as a Therapeutic Strategy for COVID-19. *ACS Infect Dis*. 2021 7:1457-1468.
277. Babin BM, Fernandez-Cuervo G, Sheng J, Green O, Ordonez AA, Turner ML, Keller LJ, Jain SK, Shabat D, **Bogyo M**. (2020) A chemiluminescent protease probe for rapid, sensitive, and inexpensive detection of live *Mycobacterium tuberculosis*. *ACS Cent Sci*. 2021 7:803-814.

278. Rompikuntal PK, Kent RS, Foe IT, Deng B, **Bogyo M**, Ward GE. Blocking Palmitoylation of *Toxoplasma gondii* Myosin Light Chain 1 Disrupts Glideosome Composition but Has Little Impact on Parasite Motility. *mSphere*. 2021 May 19;6(3):e00823-20.
279. Onguka O, Babin BM, Lakemeyer M, Foe IT, Amara N, Terrell SM, Lum KM, Cieplak P, Niphakis MJ, Long JZ, **Bogyo M**. *Toxoplasma gondii* serine hydrolases regulate parasite lipid mobilization during growth and replication within the host. *Cell Chem Biol*. 2021 May 25:S2451-9456(21)00218-X.
280. Amara N, Cooper MP, Voronkova MA, Webb BA, Lynch EM, Kollman JM, Ma T, Yu K, Lai Z, Sangaraju D, Kayagaki N, Newton K, **Bogyo M**, Staben ST, Dixit VM. Selective activation of PFKL suppresses the phagocytic oxidative burst. *Cell*. 2021 Aug 19;184(17):4480-4494.e15.
281. Bogyo M. A 'Swiss army knife' probe for metastatic cancers. *Nat Mater*. 2021 Oct;20(10):1312-1314.
282. Babin BM, Keller LJ, Pinto Y, Li VL, Eneim AS, Vance SE, Terrell SM, Bhatt AS, Long JZ, Bogyo M. Identification of covalent inhibitors that disrupt *M. tuberculosis* growth by targeting multiple serine hydrolases involved in lipid metabolism. *Cell Chem Biol*. 2021 Sep 20:S2451-9456(21)00402-5.
283. Muir RK, Guerra M, **Bogyo M**. Activity-Based Diagnostics: Recent Advances in the Development of Probes for Use with Diverse Detection Modalities. *ACS Chem Biol*. 2022 Jan 13. doi: 10.1021/acscchembio.1c00753.
284. Lakemeyer M, **Bogyo M**. Uncovering an overlooked consequence of phosphorylation: change in cysteine reactivity. *Nat Methods*. 2022 Mar;19(3):281-283. doi: 10.1038/s41592-022-01414-5.
285. Keller LJ, Lakemeyer M, **Bogyo M**. Integration of bioinformatic and chemoproteomic tools for the study of enzyme conservation in closely related bacterial species. *Methods Enzymol*. 2022;664:1-22. doi: 10.1016/bs.mie.2021.11.017. Epub 2021 Dec 23.
286. Walker E, Linders DGJ, Abenojar E, Wang X, Hazelbag HM, Straver ME, Bijlstra OD, March TL, Vahrmeijer AL, Exner A, **Bogyo M**, Basilion JP, Straight B. Formulation of a Thermosensitive Imaging Hydrogel for Topical Application and Rapid Visualization of Tumor Margins in the Surgical Cavity. *Cancers (Basel)*. 2022 Jul 16;14(14):3459. doi: 10.3390/cancers14143459.
287. Richards CM, Jabs S, Qiao W, Varanese LD, Schweizer M, Mosen PR, Riley NM, Klüssendorf M, Zengel JR, Flynn RA, Rustagi A, Widen JC, Peters CE, Ooi YS, Xie X, Shi PY, Bartenschlager R, Puschnik AS, **Bogyo M**, Bertozzi CR, Blish CA, Winter D, Nagamine CM, Braulke T, Carette JE. The human disease gene LYSET is essential for lysosomal enzyme transport and viral infection. *Science*. 2022 Oct 7;378(6615):eabn5648. doi: 10.1126/science.abn5648. Epub 2022 Oct 7.
288. Brown RWB, Sharma AI, Villanueva MR, Li X, Onguka O, Zilbermintz L, Nguyen H, Falk BA, Olson CL, Taylor JM, Epting CL, Kathayat RS, Amara N, Dickinson BC, **Bogyo M**, Engman DM. Trypanosoma brucei Acyl-Protein Thioesterase-like (TbAPT-L) Is a Lipase with Esterase Activity for Short and Medium-Chain Fatty Acids but Has No Depalmitoylation Activity. *Pathogens*. 2022 Oct 27;11(11):1245. doi: 10.3390/pathogens11111245.
289. **Bogyo M**. Finding optimal drug target sites in parasite pathogens. *Trends Parasitol*. 2023 Feb;39(2):83-85. doi: 10.1016/j.pt.2022.12.003. Epub 2022 Dec 21.
290. Faucher FF, Abegg D, Ipock P, Adibekian A, Lovell S, **Bogyo M**. Solid Phase Synthesis of Fluorosulfate Containing Macrocycles for Chemoproteomic Workflows. *Isr. J. Chem*. 2023, 63, e202300020.
291. Deni I, Stokes BH, Ward KE, Fairhurst KJ, Pasaje CFA, Yeo T, Akbar S, Park H, Muir R, Bick DS, Zhan W, Zhang H, Liu YJ, Ng CL, Kirkman LA, Almaliti J, Gould AE, Duffey M, O'Donoghue AJ,

Uhlemann AC, Niles JC, da Fonseca PCA, Gerwick WH, Lin G, **Bogyo M**, Fidock DA. Mitigating the risk of antimalarial resistance via covalent dual-subunit inhibition of the Plasmodium proteasome. *Cell Chem Biol.* 2023 May 18;30(5):470-485.e6. doi: 10.1016/j.chembiol.2023.03.002.

- 292.** Faucher FF, Liu KJ, Cosco ED, Widen JC, Sorger J, Guerra M, **Bogyo M**. Protease Activated Probes for Real-Time Ratiometric Imaging of Solid Tumors. *ACS Cent Sci.* 2023 May 4;9(5):1059-1069. doi: 10.1021/acscentsci.3c00261.
- 293.** Keller LJ, Nguyen TH, Liu LJ, Hurysz BM, Lakemeyer M, Guerra M, Gelsinger DJ, Chanin R, Ngo N, Lum KM, Faucher F, Ipock P, Niphakis MJ, Bhatt AS, O'Donoghue AJ, Huang KC, **Bogyo M**. Chemoproteomic identification of a DPP4 homolog in *Bacteroides thetaiotaomicron*. *Nat Chem Biol.* 2023 Dec 19(12):1469-1479.
- 294.** Bennett JM, Ward KE, Muir RK, Kabeche S, Yoo E, Yeo T, Lam G, Zhang H, Almaliti J, Berger G, Faucher FF, Lin G, Gerwick WH, Yeh E, Fidock DA, **Bogyo M**. Covalent Macrocyclic Proteasome Inhibitors Mitigate Resistance in *Plasmodium falciparum*. *ACS Infect Dis.* 2023 Oct 13;9(10):2036-2047.
- 295.** Jo J, Upadhyay T, Woods EC, Park KW, Pedowitz NJ, Jaworek-Korjakowska J, Wang S, Valdez TA, Fellner M, **Bogyo M**. Development of Oxadiazolone Activity-Based Probes Targeting FphE for Specific Detection of *S. aureus* Infections. *bioRxiv.* 2023 Dec 12:2023.12.11.571116 doi: 10.1101/2023.12.11.571116. Preprint.
- 296.** Bennett JM, Narwal SK, Kabeche S, Abegg D, Hackett F, Yeo T, Li VL, Muir RK, Faucher FF, Lovell S, Blackman MJ, Adibekian A, Yeh E, Fidock DA, **Bogyo M**. Mixed Alkyl/Aryl Phosphonates Identify Metabolic Serine Hydrolases as Antimalarial Targets. *bioRxiv.* 2024 Jan 11:2024.01.11.575224. doi: 10.1101/2024.01.11.575224. Preprint.

Book Chapters

- 1. Bogyo M.** and Wang E. (2002) Proteasome Inhibitors: Complex Tools for a Complex Enzyme. *The Proteasome-Ubiquitin Proteoin Degradation Pathway*. Springer Publishing. Eds. Zwickl P and Baumeister W. p. 185- 208.
- 2. Dive V, Paulick MG, McIntyre JO, Matrisian LM and Bogyo M** (2008). Activity-Based Imaging and Biochemical Profiling Tools for Analysis of the Cancer Degradome. *The Cancer Degradome- Proteases and Cancer Biology*. Springer Publishing. Eds. Edwards DR, Hoer-Hansen G, Blasi F, Sloane BF. p. 101-135.
- 3. Edgington LE and Bogyo M.** (2011) Applications for Activity-based Probes in Drug Discovery. Royal Society of Chemistry. Ed. Bunage M. P. 33-63.
- 4. Ward G and Bogyo M.** (2014) *Toxoplasma gondii* Chemical Biology. *Toxoplasma gondii, The model Apicomplexan: Perspectives and Methods*. Elsevier. Eds. Weiss LM and Kim K. p 707-730.

Current Funding

ACTIVE

R01 EB026285

Bogyo (PI)

04/01/2018 – 01/31/25

Generation of highly selective activity-based probes using chemically modified phage

Develop chemical probes that are highly selective for protease targets using phage screening methods. This approach uses phage that display bicyclic peptides that are chemically modified with active site labels to identify highly selective covalent probes for imaging applications.

R01 EB028628

Bogyo (PI)

10/01/2019 – 09/30/24

National Institutes of Health

Dual orthogonal fluorescent protease sensors for image guided surgery

Major Goals: The development of new fluorescent imaging probes and applications for robotic surgery of tumors. This will include synthesis and testing probes in mouse models of cancer and building of a camera

system that can be used with the surgical robot to detect dye labeled probes.

R21 AI161061

12/01/2021 – 11/30/2024

Bogyo (PI)

Covalent inhibitors of host cell entry by SARS-CoV-2 for treatment of COVID-19

Major Goals: Develop cyclic peptides that covalently bind and block function of the SARS-CoV2 spike protein using phage screening methods. This approach uses phage that display cyclic peptides that are chemically modified with active site labels to identify highly selective covalent inhibitors of SARS-CoV2 infection.

R01 DK130293-01A1

(Bogyo, PI)

09/21/2021 – 07/31/2026

National Institutes of Health

Targeting bacterial proteases involved in PAR signaling to treat inflammatory bowel diseases

Major Goals: This proposal is focused on screening culture extracts from commensal gut bacteria to identify secreted proteases that alter host cell signaling through PARs to alter the pathogenesis of inflammatory bowel diseases.

U19 AI171399

(Chodera, PI)

05/01/2022 – 04/30/2025

National Institutes of Health

AI-driven Structurally enabled Antiviral Platform (ASAP)

Major Goals: This is a multi-project grant and the goals are to: 1) help develop biological assays to validate novel antiviral drug targets with a reduced potential to induce drug resistance and 2) to help develop small molecule covalent probes for the evaluation of target binding and selectivity of active fragments and advanced lead molecules. In addition, Dr. Bogyo will work to facilitate communication with the co-investigators and research personnel at Stanford with those at other sites.

U19 AI171421

(Glenn, PI)

05/01/2022 – 04/30/2025

National Institutes of Health

Development of outpatient antiviral cocktails against SARS-CoV-2 and other potential pandemic RNA viruses

Major Goals: Dr. Bogyo has a role as a co-investigator in project 5 to develop new classes of inhibitors of VEEV and CHIKV proteases for therapeutic applications. This will involve synthesizing inhibitors and testing them in enzyme assays against purified protease.

PR210545P1

(Bogyo, Fidock PIs)

07/01/2022 – 06/30/2024

Department of Defense

Targeting the Plasmodium proteasome for prophylaxis and treatment of drug-resistant malaria in US Military personnel

Major Goals: This multi-PI project aims to develop new classes of Plasmodium falciparum proteasome inhibitors for the treatment of malaria. The project involves collaborations with multiple research groups to develop three primary compound series and determine their efficacy at clearing both blood and liver stage parasites. It also aims to define mechanisms of resistance to this class of anti-malarial agents.

Overlap: none